

## Annexes

### Annex I. UNEA Resolution 1/6 Marine plastic debris and microplastics

The United Nations Environment Assembly,

*Recalling* the concern reflected in the outcome document of the United Nations Conference on Sustainable Development, entitled: “The Future We Want”, that the health of oceans and marine biodiversity are negatively affected by marine pollution, including marine debris, especially plastic, persistent organic pollutants, heavy metals and nitrogen-based compounds, from numerous marine and land-based sources, and the commitment to take action to significantly reduce the incidence and impacts of such pollution on marine ecosystems,

*Noting* the international action being taken to promote the sound management of chemicals throughout their life cycle and waste in ways that lead to the prevention and minimization of significant adverse effects on human health and the environment,

*Recalling* the Manila Declaration on Furthering the Implementation of the Global Programme of Action for the Protection of the Marine Environment from Land-based Activities adopted by the Third Intergovernmental Review Meeting on the Implementation of the Global Programme of Action for the Protection of the Marine Environment from Land-based Activities, which highlighted the relevance of the Honolulu Strategy and the Honolulu Commitment and recommended the establishment of a global partnership on marine litter,

*Taking note* of the decisions adopted by the eleventh Conference of the Parties to the Convention on Biological Diversity on addressing the impacts of marine debris on marine and coastal biodiversity,

*Recalling* that the General Assembly declared 2014 the International Year of Small Island Developing States and that such States have identified waste management among their priorities for action,

*Noting with concern* the serious impact which marine litter, including plastics stemming from land and sea-based sources, can have on the marine environment, marine ecosystem services, marine natural resources, fisheries, tourism and the economy, as well as the potential risks to human health;

1. Stresses the importance of the precautionary approach, according to which lack of full scientific certainty should not be used for postponing cost-effective measures to prevent environmental degradation, where there are threats of serious or irreversible damage;
2. Recognizes the significant risks arising from the inadequate management and disposal of plastic and the need to take action;
3. Encourages governments, intergovernmental organizations, non-governmental organizations, industry and other relevant actors to cooperate with the Global Partnership on Marine Litter in its implementation of the Honolulu Strategy and to facilitate information exchange through the online marine litter network;
4. Recognizes that plastics, including microplastics, in the marine environment are a rapidly increasing problem due to their large and still increasing use combined with the inadequate management and disposal of plastic waste, and because plastic debris in the marine environment is steadily fragmenting into secondary microplastics;
5. Also recognizes the need for more knowledge and research on the source and fate of microplastics and their impact on biodiversity, marine ecosystems and human health, noting recent knowledge that such particles can be ingested by biota and could be transferred to higher levels in the marine food chain, causing adverse effects;
6. Notes that microplastics may also contribute to the transfer in the marine ecosystems of persistent organic pollutants, other persistent, bioaccumulative and toxic substances and other contaminants which are in or adhere to the particles;
7. Recognizes that microplastics in the marine environment originate from a wide range of sources, including the breakdown of plastic debris in the oceans, industrial emissions and sewage and run-off from the use of products

containing microplastics;

8. Emphasizes that further urgent action is needed to address the challenges posed by marine plastic debris and microplastics, by addressing such materials at source, by reducing pollution through improved waste management practices and by cleaning up existing debris and litter;

9. Welcomes the establishment of the Global Partnership on Marine Litter launched in Rio de Janeiro, Brazil, in June 2012 and the convening of the first Partnership Forum in 2013;

10. Also welcomes the adoption by the contracting parties to the Convention for the Protection of the Marine Environment and the Coastal Region of the Mediterranean (Barcelona Convention) at its eighteenth ordinary meeting, held in Istanbul, Turkey, from 3 to 6 December 2013, of the Regional Action Plan on Marine Litter Management, the world's first such action plan, and welcomes the draft Action Plan on Marine Litter for the North-East Atlantic region awaiting adoption by the Commission of the Convention for the Protection of the Marine Environment of the North-East Atlantic at its meeting in Cascais, Portugal, and encourages governments to collaborate through relevant regional seas conventions and river commissions with a view to adopting such action plans in their regions;

11. Requests the Executive Director to support countries, upon their request, in the development and implementation of national or regional action plans to reduce marine litter;

12. Welcomes the initiative by the Joint Group of Experts on the Scientific Aspects of Marine Environmental Protection to produce an assessment report on microplastics, which is scheduled to be launched in November 2014;

13. Also welcomes the work undertaken by the International Whaling Commission on assessing the impacts of marine debris on cetaceans and the endorsement by the Conference of the Parties to the Convention on the Conservation of Migratory Species of Wild Animals at its tenth meeting of resolution 10.4, addressing the impacts of marine debris on migratory species;

14. Requests the Executive Director, in consultation with other relevant institutions and stakeholders, to undertake a study on marine plastic debris and marine microplastics, building on existing work and taking into account the most up-to-date studies and data, focusing on:

(a) Identification of the key sources of marine plastic debris and microplastics;

(b) Identification of possible measures and best available techniques and environmental practices to prevent the accumulation and minimize the level of microplastics in the marine environment;

(c) Recommendations for the most urgent actions;

(d) Specification of areas especially in need of more research, including key impacts on the environment and on human health;

(e) Any other relevant priority areas identified in the assessment of the Joint Group of Experts on the Scientific Aspects of Marine Environmental Protection;

15. Invites the secretariats of the Stockholm Convention on Persistent Organic Pollutants, the Basel Convention on the Control of Transboundary Movements of Hazardous Wastes and their Disposal and relevant organizations involved in pollution control and chemicals and waste management and the secretariats of the Convention on Biological Diversity, the Convention on Migratory Species and the regional seas conventions and action plans to contribute to the study described in paragraph 14 of the present resolution;

16. Encourages governments and the private sector to promote the more resource-efficient use and sound management of plastics and microplastics;

17. Also encourages governments to take comprehensive action to address the marine plastic debris and microplastic issue through, where appropriate, legislation, enforcement of international agreements, provision of adequate reception facilities for ship-generated wastes, improvement of waste management practices and support for beach clean-up activities, as well as information, education and public awareness programmes;

18. Invites governments, intergovernmental organizations, the scientific community, non-governmental organizations, the private sector and other stakeholders to share relevant information with the Executive Director pertinent to the study described in paragraph 14;

19. Invites those in a position to do so to provide financial and other support to conduct the study identified in paragraph 14;

20. Requests the Executive Director to present the study on microplastics for the consideration of the United Nations Environment Assembly at its second session.

Annex II. a) Common chemical additives in plastics; b) Common organic contaminants absorbed by plastics

**a) Common chemical additives in plastics**

<b>Short form</b>	<b>Full name</b>	<b>Examples of function</b>
BPA	Bisphenyl A	A monomer used in the manufacture of polycarbonates and epoxy resins
DBP	Dibutyl Phthalate	Anti-cracking agents in nail varnish
DEP	Diethyl Phthalate	Skin softeners, colour and fragrance fixers
DEHP	Di-(2-ethylhexyl)phthalate	Plasticizer in PVC
HBCD	Hexabromocyclododecane	Flame retardant
PBDEs	Polybrominated Diphenyl Ethers (penta, octa & deca forms)	Flame retardants
	Nonylphenol	Stabilizer in PP, PS
phthalates	Phthalate esters	Improve flexibility and durability

**b) Common organic contaminants absorbed by plastics**

<b>Short form</b>	<b>Full name</b>	<b>Origin</b>
DDT	Dichlorodiphenyltrichloroethane	Insecticide
PAHs	Polycyclic Aromatic Hydrocarbons	Combustion products
PCBs	Polychlorinated Biphenyls	Cooling and insulating fluids, e.g. in transformers

Annex III. Examples of concentrations of plastics in rivers flowing to the ocean

Table III.1. Examples of concentrations of plastics in rivers flowing to the ocean (adapted from GESAMP 2016)

Location	Compartment	Sampling	Abundance (densities)	References
<b>Europe</b>				
Danube river, Austria, Europe	Surface water	Sizes classes: <2mm, 2-20mm Sampling mesh: 500mm	Max: 141 647.7 items /1000m <sup>-3</sup> Mean: 316.8 (±4664.6) items/ 1000 m <sup>3</sup> 73.9% represent spherules (~3mm)	Lechner 2014
Elbe, Mosel, Neckar and Rhine rivers, Germany, Europe	Sediment	Size classes: <5mm	Max: 64 items kg <sup>-1</sup> dry weight, Mean: not indicated	Wagner 2014
Po river/Adriatic Sea	Surface water	Neuston net (330µm), Monthly,	1 (Spring) to 12.2 items m <sup>-3</sup> (winter)	Vianello 2013
Seine river/ English Channel	Surface water	A plankton net (80mm mesh), and (ii) a manta trawl (330mm mesh)	(i) Plankton net: 3-108 particles/m <sup>3</sup> . (ii) Manta trawl: 0.28–0.47particles m <sup>-3</sup>	Dris 2015
Rhine, Main Rivers, Germany	Sediment	63–5000µm Three size classes: 630–5000, 200–630, and 63–200µm	Range: 228–3763 particles kg <sup>-1</sup>	Klein 2015
Solent: Hamble, Itchen and Test as tributaries to Southampton Water in Hampshire, UK	Surface water	1235 (total of 4 samples) sampled in each estuary. 0.3mm mesh	Itchen 1.55mp m <sup>-2</sup> Test 5.86m <sup>-2</sup> Hamble 0.4mp m <sup>-2</sup>  Total all estuaries: 3.72m <sup>-2</sup> (Southampton water 1.29m <sup>-2</sup> )	Gallagher 2015
Tamar estuary, UK, Europe	Surface water	Size classes: <1mm, 1e3mm, 3e5mm, >5mm Sampling mesh: 300mm	Max: 204 pieces of suspected plastic Mean: 0.028 items m <sup>3</sup> Abundances include all plastic particles, of which 82% represents size <5mm	Sadri 2014
<b>North America</b>				
North Shore Channel (Chicago, USA)	Surface water	Two neuston nets (0.92 × 0.42m and 0.36 × 0.41m) of 333-µm mesh	Upstream waters : 1.94 (0.81) particles m <sup>-3</sup> Downstream waters : 17.93 (11.05) particles m <sup>-3</sup>	McCormick 2014
St. Lawrence River, Canada/USA,	Sediment	Size classes: not indicated. Items size range: 0.4-2.16mm	Mean: 13 759 (±13 685) items m <sup>-2</sup> max at 136 926 (±83947) items m <sup>-2</sup>	Castañeda 2014
Los Angeles River, San Gabriel River, Coyote Creek, USA, North America	Surface, mid and near-bottom water	Size classes: >1.0 and <4.75mm, >4.75mm Sampling mesh: 333, 500, and 800µm	Max: 12 932 items m <sup>-3</sup> Mean 24-h particle counts on date of greatest abundance: Coyote creek: 5000 items m <sup>-3</sup> San Gabriel river: 51 603 items m <sup>-3</sup> Los Angeles River: 1 146 418 items m <sup>-3</sup> Item size class: 1.0-4.75mm	Moore 2011
<b>South America</b>				
Elqui, Maipo, Maule and BioBio rivers, northern-central (29° S) to southern central Chile (37° S)	Surface water	Neuston net with a mesh size of 1mm and an opening area of 27 * 10.5cm <sup>2</sup> . 6 2 counts by scientists + 2-6 counts by students	Elqui Mouth : 0.12875m <sup>-3</sup> Maipo: 0.647m <sup>-3</sup> Maule: 0.74m <sup>-3</sup> BioBio: 0.05m <sup>-3</sup>	Rech 2015
<b>Asia</b>				
Nakdong River (187.1 m3/s)/ Jinhae Bay, southern Korea.	Surface water	Trapping of surface water, 2mm mesh screen, 100 times, 3.14m <sup>2</sup> /2.2-2.8l. samples/station	120 particles l <sup>-1</sup> (10% paints) , 187±207 particles l <sup>-1</sup> after heavy rain	Song 2015
Yangtze Estuary, China	Surface	Pumping/filtration (32-µm steel sieve)	4137 ± 2462m <sup>-3</sup>	Zhao 2014

Annex IV. Abundance of microplastics in subtidal sediments worldwide

Table IV.1 Abundance of microplastics in subtidal sediments worldwide. Location and location specification (Modified from Van Cauwenberghe et al. 2015).

Continent	Location	Location Specification	Depth	Particle Size	Measured Abundance	Reference
Americas	US	Maine Subtidal		0.250mm-4mm	105 items/L	Graham & Thompson 2009
	US	Florida Subtidal		0.250mm-4mm	116-215 items/L	Graham & Thompson 2009
	Brazil	Tidal Plain		1mm-10cm	6.36-15.89 items/m <sup>2</sup>	Costa et al. 2011
Asia	India	Shipbreaking Yard		1.6mm-5mm	81.4mg/kg	Reddy et al. 2006
	Singapore	Mangrove		1.6mm-5mm	36.8 items/kg dry	Nor & Obbard 2014
Europe	UK	Estuary			2.4-5,6 fibres/50ml	Thompson et al. 2004
	Sweden	Subtidal		2mm-5mm	2-332 items/100ml	Noren 2007
	Belgium	Harbour		0.38mm-1mm	166.7 items/kg dry	Claessens et al. 2011
		Continental Shelf	0-200m		97.2 items/kg dry	
	Italy	Subtidal		0.7mm-1mm	672-2175 items/kg dry	Vianello et al. 2013
	Slovenia	Shelf	Infralittoral (<50m)		30-800items/kg dry	Bajt <i>et al.</i> 2015
Oceanic Sediments	Polar Ocean, Mediterranean, North Atlantic, Gulf of Guinea	Deep Sea	1176-4848	5 mm-1mm	0.5 items/cm <sup>2</sup>	Van Cauwenberghe et al. 2013
	NW Pacific	Deep Sea Trench	4869- 5766	0.300mm-5 mm	60-2020 items/m <sup>2</sup>	Fisher et al. 2015
	Subpolar/North Atlantic	Deep Sea Mount Slope	1000-2000	0.032-5mm	10-15 pieces per 50ml	Woodall et al. 2015
	North East Atlantic	Canyons/Slope	1400-2200	0.032-5mm	6-40 pieces per 50ml	Woodall et al. 2015
	Mediterranean	Canyons/Slope/Basin	300-3500	0.032-5mm	10-35 pieces per 50ml	Woodall et al. 2015
	SW Indian	Seamount	500-1000	0.032-5mm	Up to 4 pieces per 50ml	Woodall et al. 2015

## Annex V Entanglement of Cetaceans and Pinnipeds

Table V.1: Overview of literature containing data on entanglement of cetaceans (from Butterworth et al. 2012)

Species / Subspecies	Region (FAO statistical areas [FAO 2012])	Entanglement rate (% entangled each year)	Entanglement rate (by animal or by % of population with scars)	Fishing pot gear debris (%)	Net (derelict) debris (%)	Mortality estimate (%)*	Source
Humpback whale	Western Central Atlantic			41	50	10	Johnson et al. 2005
Humpback whale	North West Atlantic	2.4	17 whales become entangled each year			26	Cole et al. 2006
Humpback whale	North West Atlantic	8-10.4	48-57				Robbins & Mattila 2004
Humpback whale	North East Pacific	8	52-78				Neilson et al. 2007
Western grey whale	North West Pacific		18.7				Bradford et al. 2009
Minke whale	North East Atlantic		5-22				Northridge et al. 2010
Minke whale	North West Pacific			31	69	0.9	Song et al. 2010
Minke whale	North West Atlantic	2.6	7 whales per year			37	Cole et al. 2006
North Atlantic right whale	North West Atlantic		57	25	67	12	Kraus 1990
North Atlantic right whale	North & Central West Atlantic	1.6	6 whales per year			27	Cole et al. 2006
North Atlantic right whale	North & Central West Atlantic	1.15		71	14	29	Johnson et al. 2005
Fin whale	North East Atlantic		5				Sadove & Morreale 1990
Fin whale	North West Atlantic	0.8	2 whales per year			44	Cole et al. 2006
Blue whale	North West Atlantic		<1 whale per year				Cole et al. 2006
Bryde's whale	North West Atlantic	0.2	<1 whale per year				Cole et al. 2006

Table V.2: Overview of literature containing data on the entanglement of pinnipeds (from Butterworth et al. 2012).

Species / Subspecies	Region (FAO statistical areas [FAO 2012])	Entanglement rate (% incidence in population)	Plastic debris (%)	Net debris (%)	Fishing Line debris (%)	Mortality estimate (%)*	Source
Kalikoura fur seal	South West Pacific	0.6-2.8	31	42			Boren et al. 2006
Australian fur seal	Eastern Indian Ocean	1.9	30	40		73	Pemberton et al. 1992
New Zealand fur seal	Eastern Indian Ocean	0.9	30	29	3	57	Page et al. 2004
Australian sea lion	Eastern Indian Ocean	1.3	11	66	6	44	Page et al. 2004
Antarctic & Sub – Antarctic fur seal	Western Indian Ocean	0.24	41	17	c. 10		Hofmeyr et al. 2002
Antarctic fur seal	South East Atlantic	0.024-0.059	18	48		50	Hofmeyr et al. 2006
Antarctic fur seal	South West Atlantic	0.4	46-52			80	Arnould and Croxhall 1995
Cape fur seal	South East Atlantic	0.1-0.6	50				Shaughnessy 1980
Californian sea lion	Eastern Central Pacific	3.9-7.9		50	33		Harcourt et al. 1994
Hawaiian monk seal	Eastern Central Pacific	0.7	8	32	28	16	Henderson 2001
Stellar sea lion	North East Pacific	0.26	54	7	2		Raum-Sayuran et al. 2009
Californian sea lion	Eastern Central Pacific	0.08-0.22	25	19	14		Stewart & Yochem 1987
Northern elephant seal	Eastern Central Pacific	0.15	36	19	33		Stewart & Yochem 1987
Harbour seal	Eastern Central Pacific	0.09	33				Stewart & Yochem 1987
Northern fur seal	North East Pacific	0.24		50			Stewart & Yochem 1987



Annex VI Ingestion of microplastics by marine organisms

Table VI.1. Laboratory studies exposing organisms to microplastics. Organisms which have a commercial interest have a \* after the species name. Table includes all published studies until 11<sup>th</sup> November 2015. (Rochman et al highlighted as it is a freshwater study)

Species	Common Name	Size of Ingested Material	Polymer	Exposure Concentration	Length of exposure	Particle endpoint	Effect	Source
<b>Phylum Dinoflagellata</b>								
<i>Oxyrrhis Marina</i>		7.3µm	PS	3000 per ml	1 hr	Digestive tract	Ingestion	Cole et al. 2013
<b>Phylum Chlorophyta</b>								
<i>Tetraselmis Chuii</i>		1 – 5µm	PE	0.000046 - 0.0015 per ml	96 hrs	Cellular	No significant effect on growth, did not interact with toxicity of copper	Davarpanah & Guilhermino 2015
<i>Scenedesmus Spp.</i>		20nm	PS	1.6-40mg per ml	2 hrs	Cellular	Absorption, ROS increased, photosynthesis affected	Bhattacharya et al. 2010
<b>Phylum Haptophyta</b>								
<i>Isochrysis Galbana</i>		2µm	PS	9 x 10 <sup>4</sup> per ml	6 hrs	External	Microspheres attached to algae, no negative effect observed	Long et al. 2014
<b>Phylum Dinophyta</b>								
<i>Heterocapsa Triquetra</i>		2µm	PS	9 x 10 <sup>4</sup> per ml	6 hrs	External	Microspheres attached to algae, no negative effect observed	Long et al. 2014
<b>Phylum Cryptophyta</b>								
<i>Rhodomonas Salina</i>		2µm	PS	9 x 10 <sup>4</sup> per ml	6 hrs	External	Microspheres attached to algae, no negative effect observed	Long et al. 2014
<b>Phylum Ochrophyta</b>								
<i>Chaetoceros Neogracilis</i>		2µm	PS	9 x 10 <sup>4</sup> per ml	6 hrs	External	Microspheres attached to algae, no negative effect observed	Long et al. 2014
<b>Phylum Ciliophora</b>								
<i>Strombidium Sulcatum</i>		0.41 -10µm	-	5-10% ambient bacteria concentration	1 hr	Digestive tract	Ingestion	Christaki et al. 1998
<i>Tintinnopsis Lobiancoi</i>		10µm	PS	1000, 2000, 10000 per ml	3 hrs	Digestive tract	Ingestion	Setälä et al. 2014
<b>Phylum Cnideria</b>								
<i>Obelia Sp.</i>		20.6	PS	2240 per ml	1 hr	Digestive tract	Partial ingestion	Cole et al. 2013

<i>Dipsastrea Pallida</i>	Coral	10µm-2mm	PP	0.395mg per ml	48 hrs	Mouth and mesenteries of polyps	Ingestion	Hall et al. 2015
<b>Phylum Rotifera</b>								
<i>Synchaeta Spp.</i>		10µm	PS	2000 per ml	3 hrs	Digestive tract	Ingestion	Setälä et al. 2014
<b>Phylum Annelida</b>								
<i>Arenicola Marina</i>	Lugworm	20-2000µm	-	1.5mg per ml	Several days	Digestive tract	Ingestion	Thompson et al. 2004
<i>Arenicola Marina</i>	Lugworm	130µm	U-PVC	0-5% by weight	48 hour, 4 weeks	Digestive tract	Ingestion, reduced feeding, increased phagocytic activity, reduced available energy reserves, lower lipid reserves	Wright et al. 2013
<i>Arenicola Marina</i>	Lugworm	230µm	PVC	1500g of sediment	10 days	Digestive tract	Ingestion, oxidative stress	Browne et al. 2013
<i>Arenicola Marina</i>	Lugworm	< 5mm	HDPE, PVA, PA	0.02, 0.2 2% of sediment	31 days	Digestive tract	Concentration in sediment had significant effects on the metabolic rate of lugworms (increase mp = increase metabolic rate)	Green et al. 2015
<i>Arenicola Marina</i>	Lugworm	400-1300µm	PS	0, 1, 10, 100 mg per ml	28 days	Faeces	Ingestion, reduced feeding, weight loss	Besseling et al. 2013
<i>Galeolaria Caespitosa</i>	Fan worm	3 – 10µm	-	5000 per ml	20 mins	Digestive tract	Ingestion	Bolton & Havenhand 1998
<i>Marenzelleria Spp.</i>		10µm	PS	2000 per ml	3 hrs	Digestive tract	Ingestion	Setälä et al. 2014
<b>Phylum Mollusca</b>								
Bivalvia (larvae)		7.3µm	PS	3000 per ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
<i>Mytilus Edulis</i> *	Blue mussel	30nm	PS	0, 0.1, 0.2, 0.3 mg per ml	8 hrs	Digestive tract	Ingestion, pseudofaeces, reduced filtering	Wegner et al. 2012.
<i>Mytilus Edulis</i> *	Blue mussel	0 – 80µm	HDPE	2.5mg per ml	< 96 hrs	Digestive tract, Lymph system	Ingestion, retention in digestive tract, transfer to lymph system, immune response	Von Moos et al. 2012 & Köhler 2010
<i>Mytilus Edulis</i> *	Blue mussel	0.5µm	PS	50µL per 400 ml seawater	1 hr	Digestive tract	Ingestion, trophic transfer to <i>Carcinus maenas</i>	Farrell & Nelson 2013
<i>Mytilus Edulis</i> *	Blue mussel	3, 9.6µm	PS	0.51mg per ml	12 hrs	Digestive tract, Lymph system	Ingestion, retention in digestive tract, transferred to lymph system	Browne et al. 2008
<i>Mytilus Edulis</i> *	Blue mussel	10µm	PS	2 x 10 <sup>4</sup> per ml	45 mins	Faeces	Ingestion, egestion	Ward & Tagart 1989

<i>Mytilus Edulis</i> *	Blue mussel	10µm	PS	1000 per ml	45 mins	Faeces	Ingestion, egestion	Ward & Kach 2009
<i>Mytilus Galloprovincialis</i> *	Mediterranean mussel	< 100µm	PS, PE	1.5mg per ml	7 days	Gills, digestive tract and lymph system	presence in haemolymph, gills and digestive gland	Avio et al. 2015
<i>Mytilus Galloprovincialis</i> *	Mediterranean mussel	50nm	PS	1, 5, 50 µg per ml	-	Haemocytes	Only the haemocytes were exposed, signs of cytotoxicity	Canesi et al. 2015
<i>Mytilus Trossulus</i> *	Bay mussel	10µm	PS	/	0.5 - 1.5 hr	Digestive tract	Ingestion	Ward et al. 2003
<i>Placopecten Magellanicus</i> *	Atlantic Sea scallop	15, 10, 16, 18, 20µm	PS	1.05 per ml	1 hr	Faeces	Ingestion, retention, egestion	Brilliant & MacDonald 2000
<i>Placopecten Magellanicus</i> *	Atlantic Sea scallop	15, 10, 16, 18, 20µm	PS	1.05 per ml	1 hr	Faeces	Ingestion, retention, egestion	Brilliant & MacDonald 2002
<i>Crassostrea Virginica</i> *	Eastern oyster	10µm	PS	1000 per ml	45 mins	Faeces	Ingestion, egestion	Ward & Kach 2009
<i>Crassostrea Gigas</i> *	Pacific oyster	2, 6µm	PS	1800 per ml for the 2µm size; 200 per ml for the 6µm size	2 months	Digestive tract	Increased filtration and assimilation, reduced gamete quality, slower larval rearing for larvae from MP exposed parents	Sussarellu et al. 2014
<b>Phylum Echinodermata</b>								
<i>Apostichopus Californicus</i>	Giant Californian sea cucumber	10, 20µm	PS	2.4 per µL	-	Digestive tract	Ingestion, retention	Hart 1991
<i>Thyonella Gemmate</i>	Striped sea cucumber	0.25-15mm	PVC, PA	11g PVC shavings, 60g resin pellets, 2g nylon line, to 600ml of silica sand	20-25 hrs	Digestive tract	Selective ingestion	Graham & Thompson 2009
<i>Holothuria (Halodeima) Grisea</i>	Grey sea cucumber	0.25-15mm	PVC, PA	As above	20-25 hrs	Digestive tract	Selective ingestion	Graham & Thompson 2009
<i>Holothuria Foridana</i>	Florida sea cucumber	0.25-15mm	PVC, PA	As above	20-25 hrs	Digestive tract	Selective ingestion	Graham & Thompson 2009
<i>Cucumaria Frondosa</i> *	Orange footed sea cucumber	0.25-15mm	PVC, PA	As above	20-25 hrs	Digestive tract	Selective ingestion	Graham & Thompson 2009
<i>Paracentrotus Lividus</i> *	Sea urchin	40nm	PS	<25µg per ml	48 hr	Digestive tract	Accumulation and embryo toxicity	Della Torre et al. 2014
<i>Lytechinus Variegatus</i>	Green sea urchin	3-5mm	PE	2ml per 8ml	24 hr	External	Toxic effects, inc. anomalous embryonic development	Nombre et al. 2015
<i>Tripneustes Gratilla</i> *	Collector urchin	32-35µm	PE	1, 10, 100, 300 per ml	1-6hrs, 9 days	Faeces	Ingestion, egestion	Kaposi et al. 2014
<i>Dendraster Excentricus</i>	Eccentric sand dollar	10, 20 µm	PS	2.4 per µL	-	Digestive tract	Ingestion, retention	Hart 1991

<i>Strongylocentrotus Sp*</i>	Sea urchin	10, 20 µm	PS	2.4 per µL	-	Digestive tract	Ingestion, retention	Hart 1991
<i>Ophiopholis Aculeate</i>	Crevice brittle star	10, 20 µm	PS	2.4 per µL	-	Digestive tract	Ingestion, retention	Hart 1991
<i>Dermasterias Imbricate</i>	Leather star	10, 20 µm	PS	2.4 per µL	-	Digestive tract	Ingestion, retention	Hart 1991
<b>Phylum Arthropoda</b>								
<i>Semibalanus Balanoides</i>	Barnacle	20-2000 µm	-	1mg per ml	Several days	Digestive tract	Ingestion	Thompson et al. 2004
<i>Tigriopus Japonicas</i>	Copepod	0.05µm	PS	$9.1 \times 10^{11}$ per ml	24 hrs	Faeces	Ingestion, egestion, mortality, decreased fecundity	Lee et al. 2013
<i>Tigriopus Japonicas</i>	Copepod	0.5µm	PS	$9.1 \times 10^8$ per ml	24 hrs	Faeces	Ingestion, egestion, mortality, decreased fecundity	Lee et al. 2013
<i>Tigriopus Japonicas</i>	Copepod	6µm	PS	$5.25 \times 10^5$ per ml	24 hrs	Faeces	Ingestion, egestion, mortality, decreased fecundity	Lee et al. 2013
<i>Acartia (Acanthacartia) Tonsa</i>	Copepod	7-70 µm	-	3000-4000 beads per ml	15 mins	Digestive tract	Ingestion, size selection	Wilson 1973
<i>Acartia Spp.</i>	Copepod	10µm	PS	2000 per ml	3 hrs	Faeces	Ingestion	Setälä et al. 2014
<i>Acartia Clausi</i>	Copepod	7.3, 20.6, 30.6 µm	PS	635, 2240, 3000 beads per ml	24 hrs	Digestive tract	Size based selection: Ingestion at 7.3 µm , no ingestion at 20.6 µm, partial ingestion at 30.6 µm	Cole et al. 2013
<i>Eurytemora Affinis</i>	Copepod	10µm	PS	1000, 2000, 10,000 per ml	3 hrs	Faeces	Ingestion, egestion	Setälä et al. 2014
<i>Limnocalanus Macrurus</i>	Copepod	10µm	PS	1000, 2000, 10,000 per ml	3 hrs	Digestive tract	Ingestion	Setälä et al. 2014
<i>Temora Longicornis</i>	Copepod	1.7, 3.8, 7.3, 20.6, 30.6 µm	PS	635, 2240, 3000 beads per ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
<i>Temora Longicornis</i>	Copepod	20µm	PS	100 per ml	overnight	Digestive tract	Ingestion $10.7 \pm 2.5$ beads per individual	Cole et al. 2014
<i>Calanus Helgolandicus</i>	Copepod	20µm	PS	75 per ml	23 hrs	Faeces	Egestion, ingestion	Cole et al. 2015
<i>Calanus Helgolandicus</i>	Copepod	7.3, 20.6, 30.6 µm	PS	635, 2240, 3000 beads per ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
<i>Centropages Typicus</i>	Copepod	7.3, 20.6, 30.6 µm	PS	635, 2240, 3000 beads per ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
<i>Idotea Emarginata</i>	Isopod	10µm	PS	0.3-120 mg/g	3 days	Faeces	Ingestion, presence in stomach, faeces, no evidence of assimilation, no absorbance, no adverse effect on life history	Hamer et al. 2014
<i>Orchestia Gammarellus</i>	Amphipod	20 – 2000µm	-	1g per individual (n = 150)	several days	Digestive tract	Ingestion	Thompson et al. 2004

<i>Talitrus Saltator</i>	Amphipod	10 – 45µm	PE	10% weight (0.06-0.09 p/g dry food)	24 hrs	Faeces	Ingestion, egestion after 2 hours	Ugolini et al. 2013
<i>Allorchestes Compressa</i>	Amphipod	11 - 700µm	PE	0.1 per g	72 hrs	Faeces	Ingestion, egestion within 36 hours	Chua et al. 2014
<i>Neomysis Integer</i>	Shrimp	10µm	PS	2000 spheres per ml	3 hrs	Digestive tract	Ingestion	Setälä et al. 2014
<i>Mysis Relicta</i>		10µm	PS	2000 spheres per ml	3 hrs	Faeces	Ingestion, egestion	Setälä et al. 2014
<i>Carcinus Maenas*</i>	Shore crab	8 - 10µm	PS	4.0 x 10 <sup>4</sup> per l ventilation 1.0 x 10 <sup>6</sup> per g	16 hrs, 24 hrs, 21 days	Faeces	Ingestion through gills and gut, retention and excretion, no biological effects measured	Watts et al. 2014
<i>Carcinus Maenas*</i>	Shore crab	250-500µm	-	180mg per 9 cubes of feed	3 weeks	Digestive tract	Ingestion, MP presence did not affect PAH uptake	Msc thesis: Zoeter Vanpoucke Mechtild
<i>Uca Repax</i>	Fiddler crab	180-250µm	PS	108-1000mg/kg	2 months	Gills, Digestive tract, Lymph system	2 month exposure, 100% with MP found in gills, stomach, hepatopancreus. More MP exposure, more MP in crab. Not sure of effect	Brennecke et al. 2015
<i>Nephrops Norvegicus*</i>	Norway lobster	5mm	PP	10 fibres per cm <sup>3</sup> fish	24 hrs	Digestive tract	Ingestion	Murray and Cowie 2011
Porcellanidae (zoea)	Decopoda	30.6µm	PS	635 beads p/ml	24 hrs	Digestive tract	Partial Ingestion	Cole et al. 2013
Paguridae (zoea)	Decopoda	20.6µm	PS	2240 beads p/ml	24 hrs	Digestive tract	Partial Ingestion	Cole et al. 2013
Caridea (larvae)	Decopoda	20.6µm	PS	2240 beads p/ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
Barchyura (megalopa)	Decopoda	20.6µm	PS	2240 beads p/ml	24 hrs	Digestive tract	Ingestion	Cole et al. 2013
<i>Artemia Franciscana</i>	Brine shrimp	40 & 50 nm	PS	5-100 µg p/ml	48 hrs	Digestive tract	Ingestion, no mortality, possible effect on motility, some excretion	Bergami et al. 2015
<i>Nephrops Norvegicus*</i>	Norway lobster	500 - 600 µm loaded with 10 µg of PCBs	PE	150mg microplastics in gelatine food	3 weeks	Faeces	Ingestion, 100% egestion. Increase of PCB level in the tissues. Same increase for positive control. No direct effect of microplastics.	Devriese et al. in prep
<b>Phylum Chordata</b>								

<i>Doliolidae</i>	Tunicata	7.3µm	PS	3000 beads ml	1 hr	Digestive tract	Ingestion	Cole et al. 2013
<i>Pomatoschistus Microps</i>	Common goby	1 - 5 µm	PE	18.4 & 184 µg p/l	96 hrs	External	Abnormal swimming behaviour and lethargy, ACHe activity affected	Oliveria et al. 2013
<i>Pomatoschistus Microps</i>	Common goby	420 - 500 µm	PE	< 30 per fish	3 mins	Digestive tracts	Ingestion, significant decrease in predatory performance	De Sa et al. 2015
<i>Pomatoschistus Microps</i>	Common goby	1 - 5 µm	PE	0.216 mg p/l	/	Digestive tracts	The toxicological interaction between MP and Cr(VI) at conc >3.9 mg/l decreased predatory performance (67%) and caused significant inhibition of ACHe activity (<31%)	Luis et al. 2015
<i>Gadus Morhua</i> *	Atlantic cod	2, 5 mm	PE	/	/	Faeces	Ingestion, egestion, 5mm held for prolonged periods, emptying of plastics improved by food consumption additional meals.	Dos Santos & Jobling 1991
<i>Oryzias Latipes</i> *	Japanese medaka	<0.5mm	LDPE	Ground up as 10% of diet	1-2 months	Digestive tracts	Liver toxicity, pathology, hepatic stress	Rochman et al. 2013
<i>Oryzias Latipes</i> *	Japanese medaka	<0.5mm	LDPE	Ground up as 10% of diet	1-2 months	Digestive tracts	Altered gene expression, decreased choriogenin regulation in males and decreased vitellogenin and choriogenin in females	Rochman et al. 2014
<i>Dicentrarchus Labrax</i> *	Seabass (larvea)	10 - 45 µm	PE	0-105 per g incorporated with food	8dph - 26dph	Digestive tract	Ingestion, no significant increase in growth, effect on survival of larvae. Possible gastric obstruction.	Mazurais et al. 2014
<i>Halichoerus Grypus</i>	Grey seal	3mm	PE	2818 beads (99% recovery)	96 hours	Faeces	Used as a tracer for diet study	Grellier and Hammond 2006
<i>Calonectris leucomelas</i>	Streaked shearwater	3-5 mm	PE	1g of beads exposed to PCBs ~ 97ng per g	1st day exposed, studied for 42 days	Chemicals in preen oil	Ingestion, chemical transfer	Teuten et al. 2009

Table VI.2 Evidence of microplastic ingestion by field studies organisms. If mean not available, range is reported. Standard deviation is reported where possible.\* represents percentage ingestion by total number of individuals, not separated by species. \* species which are commercially important

Scientific name	Common name	Number of individuals	% with microplastic	Mean particles per individual (SD)	Range	Polymer	Type of microplastic	Size ingested (mm)	Study location	Source
<b>Phylum Mollusca</b>										
<i>Dosidicus gigas</i>	Humboldt squid	30	26.7	/	0-11	/	Nurdles	3-5mm	British Columbia, Canada	Braid et al. 2012
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	17	/	Total: 0.08 (0.09)-0.34 (0.22sd) p/g	/	/	Fibres, particles	<5mm	Tagus Estuary, Portugal	Vandermeersch et al. 2015
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	17	/	Mean: 0.11 (0.12)-0.15 (sd0.33) p/g	/	/	Fibres, particles	<5mm	Ebro Delta Coastal Embayment, Spain	Vandermeersch et al. 2015
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	5	/	Mean: 0.25 (0.26sd) p/g	/	/	Fibres, particles	<5mm	Goro, Italy	Vandermeersch et al. 2015
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	5	/	Mean: 0.04 (0.09sd) p/g	/	/	Fibres, particles	<5mm	Amposta, Ebro Delta, Spain	Vandermeersch et al. 2015
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	18	100	4.33 (2.62)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, china	Li et al. 2015
<i>Mytilus galloprovincialis</i> *	Mediterranean mussel	17	/	Mean 0.05 (0.11)-0.16 (0.11 sd) p/g	/	/	Fibres, particles	<5mm	Po estuary, Italy	Vandermeersch et al. 2015
<i>Mytilus edulis</i> *	Blue mussel	5	/	Mean 0.06 ( $\pm$ 0.13) particles p/g	/	/	Fibres	<5mm	Baie de Saint Brioux, France	Vandermeersch et al. 2015
<i>Mytilus edulis</i> *	Blue mussel	5	/	Mean 0.32 ( $\pm$ 0.22) p/g	/	/	Fibres, particles	<5mm	Inschot, The Netherlands	Vandermeersch et al. 2015
<i>Mytilus edulis</i> *	Blue mussel	45	/	3.5 per 10g	/	/	Fibres	300-1000 $\mu$ m	Belgium, The Netherlands	De Witte et al. 2014
<i>Mytilus edulis</i> *	Blue mussel	36	/	0.36 ( $\pm$ 0.07) p/g	/	/	/	5-25 $\mu$ m	North Sea, Germany	Van Cauwenberghe & Janssen 2014
<i>Mytilus edulis</i> *	Blue mussel	20	/	170-375 particles per 5 mussels	/	/	Fibres	/	Nova Scotia, Canada	Mathlon & Hill 2014
<i>Scapharca subcrenata</i> *	Ark shell	6	100	45 ( $\pm$ 14.98)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Tegillarca granosa</i> *	Blood cockle	18	100	5.33 ( $\pm$ 2.21)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015

<i>Patinopecten yessoensis</i> *	Yesso Scallop	6	100	57.17 (± 17.34)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Alectryonella plicatula</i> *	Fingerprint oyster	18	100	10.78 (± 4.07)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Sinonovacula constricta</i> *	Chinese razor clam	6	100	14.33 (± 5.35)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Ruditapes philippinarum</i> *	Carpet shell	24	100	5.72 (± 2.86)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Meretrix lusoria</i> *	Orient clam	18	100	9.22(± 0.46)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Cyclina sinensis</i> *		30	100	4.82 (± 2.17)	/	PET, PA, PE	Fibres, fragments, pellets	<5mm	Fish market, China	Li et al. 2015
<i>Crassostrea gigas</i> *	Pacific oyster	12	30	0.6±0.9	0-2	/	Fibres		USA	Rochman et al. 2015
<i>Crassostrea gigas</i> *	Pacific oyster	11	/	0.47(± 0.16) per g	/	/		5-25 µm	Atlantic Ocean	Van Cauwenberghe & Janssen 2014
<b>Phylum Arthropoda</b>										
Lepas spp. *	Gooseneck barnacle	385	33.5	/	01/30/16	/		<5mm	North Pacific	Goldstein & Goodwin 2013
<i>Neocalanus cristatus</i>	Calanoid copepod	960	/	1 particle per 34 zoop			Fibre, fragment	556 (149) µm	North Pacific	Desforges et al. 2015
<i>Euphausia pacifica</i>	Euphausiid	413	/	1 particle per 7 euph			Fibre, fragment	816 (108) µm	North Pacific	Desforges et al. 2015
<i>Nephrops norvegicus</i> *	Norway lobster	120	83	/					Clyde, UK	Murray and Cowie 2011
<i>Crangon crangon</i> *	Brown shrimp	110	/	11.5 fibres per 10 g			95% fibres, 5% films	300-1000 µm	Belgium	Devriese et al. 2015
<b>Phylum Annelida</b>										
<i>Arenicola marina</i>	Lugworm			1.2 +- 2.8 g/ w.w				>5µm	Belgium, NL, France	Van Cauwenberge et al. in Devriese et al. 2015
<b>Phylum Chaetognatha</b>										
<i>Parasagitta elegans</i>	Arrow worm	1	100	/		PS	Spheres	0.1-3mm	New England, USA	Carpenter et al. 1972
<b>Phylum Chordata</b>										
<i>Phoca vitulina</i>	Harbour seal	100 stomachs 107	S:11.2 , I:1	Max: 8 items (s), 7 items (i)			Fragments	>0.1	The Netherlands	Bravo Rebolledo et al. 2013



intestines									
<i>Mesoplodon mirus</i>	True's beaked whale	1	100	88		Fibres, fragment	mean 2.16mm	Connemara, Ireland	Lusher et al. 2015
<i>Megaptera novaeangliae</i>	Humpback whale	1	100	45 items		Fragments	1-17cm	The Netherlands	Besseling et al. 2015
<i>Arctocephalus</i> spp.	Fur seal	145	100	1-4 per scat		Fragments, beads	4.1mm	Macquarie Island, Australia	Eriksson & Burton 2003
<i>Chelonia mydas</i> *	Green turtle	24	/	Total: 11 pellets		Pellets	<5mm	Rio Grande do Sul, Brazil	Tourinho et al. 2010
<i>Menidia menidia</i>	Atlantic silversides	9	33	/		PS	0.1-3mm	New England, USA	Carpenter et al 1972
<i>Atherinopsis californiensis</i> *	Jacksmelt	7	28.5714 2857	1.6+-3.7		Fibres, fragments	0-10	USA	Rochman et al. 2015
<i>Alepisaurus ferox</i>	Longnosed lancetfish	144	24	2.7 (± 2.0)		Fragments	68.3 (±91.1)	North Pacific	Choy and Drazen 2013
<i>Cololabis saira</i>	Pacific saury	52	*35	3.2 (± 3.05)		Fragments	0--2	North Pacific	Boerger et al. 2010
<i>Clupea harengus</i> *	Atlantic herring	2	100	1	PS	PS	0.1 -3mm	New England, USA	Carpenter et al 1972
<i>Clupea harengus</i> *	Atlantic herring	566	2		1 to 4	Fragments	0.5-3	North Sea	Foekema et al. 2013
<i>Clupea harengus</i> *	Atlantic herring	3	100	/		/	/	North Sea	Collard et al. 2015
<i>Sprattus sprattus</i> *	European sprat	111	38.74%	0.88 (0.88)		Fibres, granual, film	0.1-4.9mm	Belgium, North Sea	Msc thesis: Zoeter Vanpoucke Mechtild
<i>Spratelloides gracilis</i> *	Silverstripe round herring	4	40	1.1 +-1.7	0-5	0-5 fragments		Indonesia	Rochman et al. 2015
<i>Alosa fallax</i> *	Twait shad	1	100	1		Fragment	<5mm	North Eastern Atlantic	Neves et al., 2015
<i>Sardina pilchardus</i> *	European pilchard	3	100%	/		/	/	North Sea	Collard et al. 2015
<i>Sardina pilchardus</i> *	European pilchard	99	19%	1.78 ± 0.7			<1mm	Adriatic sea	Avio et al. 2015
<i>Sardinella longicxeps</i> *	Oil sardine	10	60%	/		Fibres	0.5-3mm	Mangalore	Sulochanan et al. 2014
<i>Stolephorus commersonii</i> *	Anchovy	16	37.5	/		Fragments	1.14-2.5	Alappuzha, India	Kripa et al. 2014
<i>Engraulis encrasicolus</i> *	Anchovy	3	100%	/		/	/	North Sea	Collard et al. 2015

<i>Engraulis mordax</i> *	Pacific anchovy	10	30	0.3+-0.5	0-1	Fibres and film		USA	Rochman et al. 2015
<i>Pollachius virens</i> *	Saithe	1	100	1		PS	PS	0.1-3mm PS	New England, USA Carpenter et al 1972
<i>Ciliata mustela</i>	Five-bearded rocklings	113	0-10	/		PS	PS	2mm	Severn Estuary, UK Kartar 1976
<i>Merlangius merlangus</i> *	Whiting	105	6	01/03/16				1.7 (±1.5)	North Sea Foekema et al. 2013
<i>Merlangius merlangus</i> *	Whiting	50	32	1.75 (± 1.4)			Fragment, fibres, beads	2.2 (±2.3)	English Channel Lusher et al. 2013
<i>Melanogrammus aeglefinus</i> *	Haddock	97	6	1			Fragments	0.7 (±0.3)	North Sea Foekema et al. 2013
<i>Gadus morhua</i> *	Cod	80	13	01/02/16			Fragments	1.2 (±1.2)	North Sea Foekema et al. 2013
<i>Micromesistius poutassou</i> *	Blue whiting	27	51.9	2.07 (± 0.9)			Fragment, fibres, beads	2.0 (±2.4)	English Channel Lusher et al. 2013
<i>Trisopterus minutus</i> *	Poor cod	50	40	1.95 (± 1.2)			Fragment, fibres, beads	2.2 (±2.2)	English Channel Lusher et al. 2013
<i>Merluccius merluccius</i> *s	Hake	3	100%	1.33 ± 0.57				<1mm	Adriatic sea Avio et al. 2015
<i>Merluccius merluccius</i> *	Hake	12	25%	0.33±0.65			4 fibres	<5mm	North Eastern Atlantic Neves et al. 2015
<i>Lampris</i> sp. (big eye)		115	29	2.3 (± 1.6)			Fragments	49.1 (±71.1)	North Pacific Choy & Drazen 2013
<i>Lampris</i> sp. (small eye)		24	5	5.8 (± 3.9)			Fragments	48.8 (±34.5)	North Pacific Choy & Drazen 2013
<i>Lophius piscatorius</i> *	Monkfish	2	50	0.5			1 fibre	<5mm	North Eastern Atlantic Neves et al. 2015
<i>Hygophum reinhardtii</i>		45	*35	1.3 (± 0.71)			Fragments	1 – 2.79	North Pacific Boerger et al. 2010
<i>Loweina interrupta</i>		28	*35	1			Fragments	1 – 2.79	North Pacific Boerger et al. 2010
<i>Myctophum aurolaternatum</i>		460	*35	6.0 (± 8.99)			Fragments	1 – 2.79	North Pacific Boerger et al. 2010
<i>Symbolophorus californiensis</i>		78	*35	7.2 (± 8.39)			Fragments	1 – 2.79	North Pacific Boerger et al. 2010
<i>Diaphus anderseni</i>	Anderson's lanternfish	13	15.4	1			Fragments		North Pacific Davison & Asch 2011
<i>Diaphus fulgens</i>		7	28.6	1			Fragments		North Pacific Davison & Asch 2011
<i>Diaphus phillipsi</i>	Boluin's	1	100	1			Fragments	0.5	North Pacific Davison & Asch

	lanternfish								2011
<i>Lobianchia gemellarii</i>	Coco's lanternfish	3	33.3	1		Fragments		North Pacific	Davison & Asch 2011
<i>Myctophum nitidulum</i>	Pearly lanternfish	25	16	1.5		Fragments	5.46	North Pacific	Davison & Asch 2011
<i>Morone americana</i>	White perch	12	33	/	PS	PS	0.1-3mm	New England, USA	Carpenter et al. 1972
<i>Tautogolabrus adspersus</i>	Bergall	6	< 83	/	PS	PS	0.1-3mm	New England, USA	Carpenter et al. 1972
<i>Pomatoschistus minutus (As Gobius minutus)</i>	Goby	200	0 – 25	/	PS	PS	2mm	Severn Estuary, UK	Kartar 1976
<i>Argyrosomus regius*</i>	Meagre	5	60	0.80 (±0.8)		2 fragments, 2 fibres	<5mm	North Eastern Atlantic	Neves et al. 2015
<i>Stellifer brasiliensis</i>		330	9.2	0.33 – 0.83		Fragments	<1	Goiana Estuary, Brazil	Dantas et al. 2012
<i>Stellifer stellifer</i>		239	6.9	0.33 – 0.83		Fragments	<1	Goiana Estuary, Brazil	Dantas et al. 2012
<i>Eugerres brasilianus</i>		240	16.3	1–5		Fragments	1 – 5	Goiana Estuary, Brazil	Ramos et al. 2012
<i>Eucinostomus melanopterus</i>		141	9.2	1–5		Fragments	1 – 5	Goiana Estuary, Brazil	Ramos et al. 2012
<i>Diapterus rhombeus</i>		45	11.1	1–5		Fragments	1 – 5	Goiana Estuary, Brazil	Ramos et al. 2012
		7	71	5.+ -5.2	0-24			Indonesia	Rochman et al. 2015
<i>Trachurus trachurus*</i>	Horse mackerel	100	1	1		Fragments	2.52	North Sea	Foekema et al. 2013
<i>Trachurus trachurus*</i>	Horse mackerel	44	7	0.07±0.25		2 fragments; 1 fiber	<5mm	North Eastern Atlantic	Neves et al. 2015
<i>Trachurus trachurus*</i>	Horse mackerel	56	28.6	1.5 (± 0.7)		Fragment, fibres, beads	2.2 (±2.2)	English Channel	Lusher et al. 2013
<i>Trachurus picturatus*</i>	Blue jack mackerel	29	3.00%	0.03±0.18		1 fibre	<5mm	North Eastern Atlantic	Neves et al. 2015
<i>Seriola lalandi*</i>	Yellowtail amberjack	19	10.5	1		Fragments	0.5 – 11	North Pacific	Gassel et al. 2013
<i>Decapterus macrosoma</i>	Shortfin scad	17	29	2.5 +- 6.3	0-21	Fragments and PS		Indonesia	Rochman et al. 2015
<i>Callionymus lyra</i>	Dragonette	50	38	1.79 (± 0.9)		Fragment, fibres, beads	2.2 (±2.2)	English Channel	Lusher et al. 2013
<i>Cepola macrophthalma</i>	Red band fish	62	32.3	2.15 (± 2.0)		Fragment, fibres, beads	2.0 (±1.9)	English Channel	Lusher et al. 2013

Morone saxatilis	Striped bass	7	28.5714 2857	0.9+- 1.2	0-3	Bibre, film, foam	USA	Rochman et al. 2015
<i>Mullus barbatus</i> *	Red mullets	11	64%	1.57 ± 0.78		<1mm	Adriatic sea	Avio et al. 2015
<i>Mullus surmulletus</i> *	Striped red mullet	4	100%	1.75±0.5		7 fibers	North Eastern Atlantic	Neves et al. 2015
<i>Boops boops</i> *	Bogue	32	9	0.09 (±0.3)		1 fragment, 2 fibres	North Eastern Atlantic	Neves et al. 2015
<i>Dentex macrophthalmus</i> *	Large-eye dentex	1	100	1		1 fibre	North Eastern Atlantic	Neves et al. 2015
<i>Brama brama</i> *	Atlantic pomfret	3	33	0.67±1.2		2 fibres	North Eastern Atlantic	Neves et al. 2015
<i>Thunnus thynnus</i> *	Bluefin tuna	34	32.40%	/		>0.63mm	Mediterranean	Romeo et al.2015
<i>Thunnus alalunga</i> *	Albacore tuna	2	50.00%		PE	<3cm	Arabian Sea	Sajikumar et al. 2013
<i>Thunnus alalunga</i> *	Albacore tuna	131	12.90%	/		>3.60mm	Mediterranean	Romeo et al.2015
<i>Rastrelliger kanagurta</i> *	Indian Mackerel	10	50.00%	/		Fibres	Mangalore	Sulochanan et al. 2014
<i>Rastrelliger kanagurta</i> *	Indian Mackerel	9	56	1 (+- 1.1)	0-3	Fragments, pellets	Indonesia	Rochman et al. 2015
<i>Scomber japonicas</i> *	Chub mackerel	35	31	0.57±1.04		14 fragments; 6 fibres	<9.42mm North Eastern Atlantic	Neves et al. 2015
<i>Scomber scombrus</i> *	Atlantic mackerel	13	31	0.46±0.78		3 fragments; 3 fibres	<5mm North Eastern Atlantic	Neves et al. 2015
<i>siganus argenteus</i>	Streamlined spinefoot	2	50	0.5+-0.7		0-1 fragments	Indonesia	Rochman et al. 2015
<i>Siganus canaliculatus</i>	Rabbitfish	3	29	0.3-0.6		0-1 fragments	Indonesia	Rochman et al. 2015
<i>Xiphias gladius</i> *	Swordfish	56	12.50%	/		>3.69mm	Mediterranean	Romeo et al.2015
<i>Pagellus acarne</i> *	Axillary seabream	1	100	1		1 fiber	<5mm North Eastern Atlantic	Neves et al., 2015
<i>Citharichthys sordidus</i> *	Pacific sandab	5	60	1+-1.2	0-3	Fibre and dilm	USA	Rochman et al. 2015
<i>Pseudopleuronectes americanus</i> *	Winter Flounder	95	2.1	/	PS	PS	0.1-3mm New England, USA	Carpenter et al 1972
<i>Platichthys flesus</i> *	Flounder	/	/	/	PS	PS	1mm Severn Estuary, UK	Kartar 1973
<i>Platichthys flesus</i> *	Flounder	1090	0 – 20.7	/	PS	PS	1mm Severn Estuary, UK	Kartar 1976
<i>Buglossidium luteum</i>	Solenette	50	26	1.23 (± 0.4)		Fragment, fibres, beads	1.9 (±1.8) English Channel	Lusher et al. 2013

<i>Microchirus variegatus</i>	Thickback sole	51	23.5	1.58 ( $\pm$ 0.8)		Fragment, fibres, beads	2.2 ( $\pm$ 2.2)	English Channel	Lusher et al. 2013
<i>Oncorhynchus tshawytscha</i> *	Chinook salmon	4	25	0.25+-0.5	0-1	Fibre		USA	Rochman et al. 2015
<i>Myoxocephalus aenaeus</i>	Grubby	47	4.2	/		PS	0.1-3mm	New England, USA	Carpenter et al 1972
<i>Ophiodon elongates</i> *	Ling cod	11	9.090909091	0.1+- 0.3	0-1	0-1 film		USA	Rochman et al. 2015
<i>Liparis liparis liparis</i>	Sea snails	220	0 – 25	/		PS	1mm	Severn Estuary, UK	Kartar 1976
<i>sebastes flavidus</i> *	Yellowtail rockfish	1	33	0.3+-0.6	0-1	Fibres		USA	Rochman et al. 2015
<i>Sebastes mystinus</i> *	Blue rockfish	10	20	0.2+-0.4	0-1	Fibres		USA	Rochman et al. 2015
<i>Chelidonichthys cuculus</i> *	Red gurnard	66	51.5	1.94 ( $\pm$ 1.3)		Fragments	2.1 ( $\pm$ 2.1)	English Channel	Lusher et al. 2013
<i>Chelidonichthys lucernus</i> *	Tub Gurnard	3	0.67	1 $\pm$ 0			<1mm	Adriatic sea	Avio et al. 2015
<i>Trigla lyra</i> *	Piper gurnard	31	19	0.26 $\pm$ 0.57		1 fragment; 7 fibers	<5mm	North Eastern Atlantic	Neves et al., 2015
<i>Prionotus evolans</i>	Striped searobin	1	100	1		PS	0.1-3mm	New England, USA	Carpenter et al 1972
<i>Cathorops spixii</i>	Madamago sea catfish	60	18.3	0.47	1 – 4			Goiana Estuary, Brazil	Possatto et al. 2011
<i>Cathorops spp</i>		60	33.3	0.55	1 – 4			Goiana Estuary, Brazil	Possatto et al. 2011
<i>Sciades herzbergii</i>	Pemecoe catfish	62	17.7	0.25	1 – 4			Goiana Estuary, Brazil	Possatto et al. 2011
<i>Astronesthes indopacificus</i>		7	*35	1		Fragments	1 – 2.79	North Pacific	Boerger et al. 2010
<i>Sternoptyx diaphana</i>	Hatchetfish	4	25	1		Fragments	1.58mm	North Pacific	Davison & Asch 2011
<i>Sternoptyx pseudobscura</i>	Highlight hatchetfish	6	16.7	1		Fragments	4.75mm	North Pacific	Davison & Asch 2011
<i>Idiacanthus antrostomus</i>	Pacific black dragon	4	25	1		Fragments	0.5mm	North Pacific	Davison & Asch 2011
<i>Zeus faber</i> *	John Dory	46	47.6	2.65 ( $\pm$ 2.5)		Fragment, fibres, beads	2.2 ( $\pm$ 2.2) mm	English Channel	Lusher et al. 2013
<i>Zeus faber</i> *	John Dory	1	100	1		Fibre	<5mm	North Eastern Atlantic	Neves et al. 2015

<i>Scyliorhinus canicula</i> *	Lesser-spotted catshark	20	20	0.27 ( $\pm 0.55$ )	1 fragment; 5 fibres	<5mm	North Eastern Atlantic	Neves et al. 2015
<i>Raja asterias</i> *	Starry ray	7	43	0.57( $\pm 0.79$ )	4 fibres	<5mm	North Eastern Atlantic	Neves et al. 2015
<i>Squalus acanthias</i> *	Spiny dogfish	9	44	1.25 ( $\pm 0.5$ )		<1mm	Adriatic sea	Avio et al. 2015

Table VI.3 Evidence of microplastic ingestion by seabirds mean ( $\pm$ SD unless \* = SE).

Species	Common name	n	Percentage with plastic (%)	Mean number of particles p/ individual	Mean size ingested $\pm$ SD (min-max) (mm)	Type of plastic	Location	Source
<b>Family Procellariidae</b>								
<i>(Aphrodroma brevirostris)</i> (as <i>Pterodroma brevirostris</i> )	Kerguelen petrel	26	3.8	1		Pellets	North Island, New Zealand	Reed 1981
<i>(Aphrodroma brevirostris)</i> (as <i>Pterodroma brevirostris</i> )	Kerguelen petrel	13	8	0.2	Mass <0.0083g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Aphrodroma brevirostris</i> (as <i>Pterodroma brevirostris</i> )	Kerguelen petrel	63	22.2	/	-	Pellets	Southern Ocean	Ryan 1987
<i>Aphrodroma brevirostris</i>	Kerguelen petrel	28	7	/	3-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Calonectris diomedea</i>	Cory's shearwater	7	42.8	/		Pellets	Southern Ocean	Ryan 1987
<i>Calonectris diomedea</i>	Cory's shearwater	147	24.5	Stomach= 2 Gizzard= 3.1		Beads	North Carolina, USA	Moser & Lee 1992
<i>Calonectris diomedea</i>	Cory's shearwater	5	100	/	<10		Rio Grande do Sul, Brazil	Colabuno et al 2009
<i>Calonectris diomedea</i>	Cory's shearwater	85	83	8 ( $\pm$ 7.9)	3.9 $\pm$ 3.5		Canary Islands, Spain	Rodríguez et al. 2012
<i>Calonectris diomedea</i>	Cory's shearwater	49	96	14.6 ( $\pm$ 24.0)	2.5 $\pm$ 6.0 <sup>A</sup>		Catalan coast, Mediterranean	Codina-García et al. 2013
<i>Daption capense</i>	Cape petrel	18	83.3	/		Pellets	Southern Ocean	Ryan 1987
<i>Daption capense</i>	Cape petrel	30	33	1	5		Ardery Island, Antarctica	Van Franeker & Bell 1988
<i>Daption capense</i>	Cape petrel	105	14	/	3-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Fulmarus glacialis</i>	Northern fulmar	3	100	7.6	1-4mm	Pellets	California, USA	Baltz & Morejohn 1976
<i>Fulmarus glacialis</i>	Northern fulmar	79	92	11.9		Pellets	Netherland and Arctic colonies	Van Franeker 1985

<i>Fulmarus glacialis</i>	Northern fulmar	8	50	3.9		Pellets	St. Kilda, UK	Furness 1985
<i>Fulmarus glacialis</i>	Northern fulmar	13	92.3	10.6		Pellets	Foula, UK	Furness 1985b
<i>Fulmarus glacialis</i>	Northern fulmar	1	100	1	4mm	Pellets	Oregon, USA	Bayer & Olson 1988
<i>Fulmarus glacialis</i>	Northern fulmar	44	86.4	Stomach = 3 Gizzard = 14	-	Beads	North Carolina, USA	Moser & Lee 1992
<i>Fulmarus glacialis</i>	Northern fulmar	19	84.2	Max: 26	-	Pellets	Alaska, USA	Robards et al. 1995
<i>Fulmarus glacialis</i>	Northern fulmar	3	100	7.7	-	Pellets	Eastern North Pacific	Blight & Burger 1997
<i>Fulmarus glacialis</i>	Northern fulmar	15	36	3.6 ( $\pm$ 2.7)	7 ( $\pm$ 4.0)		Davis Strait, Canadian Arctic	Mallory et al. 2006
<i>Fulmarus glacialis</i>	Northern fulmar	1295	95	14.6 ( $\pm$ 2.0*) – 33.2( $\pm$ 3.3*)	>1.0		North Sea	Van Franeker et al. 2011
<i>Fulmarus glacialis</i>	Northern fulmar	67	92.5	36.8 ( $\pm$ 9.8*)	>0.5		Eastern North Pacific	Avery-Gomm et al. 2012
<i>Fulmarus glacialis</i>	Northern fulmar	58	79	6.0 ( $\pm$ 0.9*)	>1.0		Westfjords, Iceland	Kühn & van Franeker 2012
<i>Fulmarus glacialis</i>	Northern fulmar	176	93	26.6 ( $\pm$ 37.5)		Fragments, pellets	Nova Scotia, Canada	Bond et al. 2014
<i>Fulmarus glacialoides</i>	Antarctic fulmar	84	2	/	2-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Fulmarus glacialoides</i>	Antarctic fulmar	9	79	/	<10		Rio Grande do Sul, Brazil	Colabuno et al 2009
<i>Halobaena caerulea</i>	Blue petrel	27	100	/		Pellets	New Zealand	Reed 1981
<i>Halobaena caerulea</i>	Blue petrel	74	85.1	/		Pellets	Southern Ocean	Ryan 1987
<i>Halobaena caerulea</i>	Blue petrel	62	56	/	3-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Pachyptila</i> spp.	Prions	/	/	/		Pellets	Gough Island, South Atlantic	Bourne & Imber 1982
( <i>Pachyptila salvini</i> )	Salvin's prion	663	20	/	2.5-3.5mm	Pellets	Wellington, New Zealand	Harper & Fowler 1987
<i>Pachyptila salvini</i>		31	51.6	/		Pellets	Southern Ocean	Ryan 1987



<i>Pachyptila belcheri</i> )	Thin-billed prion	152	6.6	/	2.5-3.5mm	Pellets	Wellington, New Zealand	Harper & Fowler 1987
<i>Pachyptila belcheri</i>	Thin-billed prion	32	68.7	/		Pellets	Southern Ocean	Ryan 1987
<i>Pachyptila vittata</i>	Broad-billed prion	31	39	0.6	Max mass: 0.066g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Pachyptila vittata</i>	Broad-billed prion	310	16.5	/	2.5-3.5mm	Pellets	Wellington, New Zealand	Harper and Fowler 1987
<i>Pachyptila vittata</i>	Broad-billed prion	137	20.4	/		Pellets	Southern Ocean	Ryan 1987
<i>Pachyptila vittata</i>	Broad-billed prion	69	10	/	3-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Pachyptila vittata</i>	Broad-billed prion	149	/	1987-89 <sup>B</sup> 1.73 ± 3.58		Pellets	Southern Ocean	Ryan 2008
<i>Pachyptila vittata</i>	Broad-billed prion	86	/	1999 <sup>B</sup> 2.93 ± 3.80		Pellets	Southern Ocean	Ryan 2008
<i>Pachyptila vittata</i>	Broad-billed prion	95	/	2004 <sup>B</sup> 2.66 ± 5.34		Pellets	Southern Ocean	Ryan 2008
<i>Pachyptila desolata</i>	Antarctic prion	35	14.3	/	2.5-3.5mm	Pellets	Wellington, New Zealand	Harper and Fowler 1987
<i>Pachyptila desolata</i>	Antarctic prion	88	47.7	/		Pellets	Southern Ocean	Ryan 1987
<i>Pachyptila desolata</i>	Antarctic prion	2	100	1	6-8.1mm		Heard Island, Australia	Auman et al. 2004
<i>Pachyptila turtur</i>	Fairy prion	105	96.2	/	2.5-3.5mm	Pellets	Wellington, New Zealand	Harper and Fowler 1987
<i>Pagodroma nivea</i>	Snow petrel	363	1	/	3-6mm	Fragments, pellets	Antarctica	Ainley et al. 1990
<i>Procellaria aequinoctialis</i>	White-chinned petrel	193	/	1983-1985 <sup>B</sup> 1.66 (± 3.04)		Pellets	Southern Ocean	Ryan 1987, 2008
<i>Procellaria aequinoctialis</i>	White-chinned petrel	526	/	2005-2006 <sup>B</sup> 1.39 (± 3.25)		Pellets	Southern Ocean	Ryan 2008
<i>Procellaria aequinoctialis</i>	White-chinned petrel	41	/	/	<10		Rio Grande do Sul, Brazil	Colabuno et al. 2009

<i>Procellaria aequinoctialis</i>	White-chinned petrel	34	44	/	<10		Rio Grande do Sul, Brazil	Colabuno et al. 2010
<i>Procellaria conspicillata</i>	Spectacled petrel	3	33	/	<10		Rio Grande do Sul, Brazil	Colabuno et al. 2010
<i>Procellaria conspicillata</i>	Spectacled petrel	9	/	/	<10		Rio Grande do Sul, Brazil	Colabuno et al. 2009
<i>Pseudobulweria rostrata</i>	Tahiti petrel	121	<1	1		Fragments	Tropical, North Pacific	Spear et al. 1995
<i>Pterodroma incerta</i>	Atlantic petrel	13	8	0.1	Max mass: 0.0053g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Pterodroma incerta</i>	Atlantic petrel	20	5	/		Pellets	Southern Ocean	Ryan 1987
<i>Pterodroma macroptera</i>	Great-winged petrel	13	7.6	/		Pellets	Southern Ocean	Ryan 1987
<i>Pterodroma mollis</i>	Soft-plumaged petrel	29	20.6	/		Pellets	Southern Ocean	Ryan 1987
<i>Pterodroma mollis</i>	Soft-plumaged petrel	18	6	0.1	0.014g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Pterodroma externa</i>	Juan Fernández petrel	183	< 1	1	3-5mm	Pellets	Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma cervicalis</i>	White-necked petrel	12	8.3	5	3-4mm	Fragments	Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma pycrofti</i>	Pycroft's petrel	5	40	2.5 ( $\pm$ 0.7)	3-5mm	Fragments and pellets	Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma leucoptera</i>	White-winged petrel	110	11.8	2.2 ( $\pm$ 3.0)	2-5mm	Fragments	Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma brevipes</i>	Collared petrel	3	66.7	1	2-5mm		Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma nigripenni</i>	Black-winged petrel	66	4.5	3.0 ( $\pm$ 3.5)	3-5mm	Fragments	Offshore, North Pacific	Spear et al. 1995
<i>Pterodroma longirostris</i>	Stejneger's petrel	46	73.9	6.8 ( $\pm$ 8.6)	2-5mm	Fragments and pellets	Offshore, North Pacific	Spear et al. 1995

<i>Puffinus Ilherminieri</i>	Audubon's shearwater	119	5	Stomach = 1 Gizzard = 4.4		Beads	North Carolina, USA	Moser & Lee 1992
<i>Puffinus assimilis</i>	Little shearwater	13	8	0.8	Max mass: 0.12g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Puffinus bulleri</i>	Buller's shearwater	3	100	8.5 ( $\pm$ 8.6)	2-8mm	Fragments and pellets	Tropical, North Pacific	Spear et al. 1995
<i>Puffinus creatopus</i>	Pink-footed shearwater	5	20	2.2	1-4mm	Pellets	California, USA	Baltz and Morejohn 1976
<i>Puffinus gravis</i>	Great shearwater	24	100	/		Beads	Briar Island, Nova Scotia	Brown et al. 1981
<i>Puffinus gravis</i>	Great shearwater	13	85	12.2	Max mass: 1.13g	Pellets	Gough Island, South Atlantic	Furness 1985a
<i>Puffinus gravis</i>	Great shearwater	55	63.6	Stomach = 1 Gizzard = 13		Beads	North Carolina, USA	Moser and Lee 1992
<i>Puffinus gravis</i>	Great shearwater	50	66	1983-1985 <sup>B</sup> 16.5( $\pm$ 19.0)		Pellets	Southern Ocean	Ryan 1987, 2008
<i>Puffinus gravis</i>	Great shearwater	53	/	2005-2006 <sup>B</sup> 11.8 ( $\pm$ 18.9)		Pellets	Southern Ocean	Ryan 2008
<i>Puffinus gravis</i>	Great shearwater	19	89	/	<10 mm		Rio Grande do Sul, Brazil	Colabuno et al. 2009
<i>Puffinus gravis</i>	Great shearwater	6	100	/	<3.2-5.3mm	Pellets	Rio Grande do Sul, Brazil	Colabuno et al. 2010
<i>Puffinus gravis</i>	Great shearwater	84	88	11.8 ( $\pm$ 16.9)		Fragments and pellets	Nova Scotia, Canada	Bond et al. 2014
<i>Puffinus griseus</i>	Sooty shearwater	21	43	5.05	1-4mm	Pellets	California, USA	Baltz and Morejohn 1976
<i>Puffinus griseus</i>	Sooty shearwater	5	100	/	Beads	Beads	Briar Island, Nova Scotia, Canada	Brown et al. 1981
<i>Puffinus griseus</i>	Sooty shearwater	36	58.3	11.4 ( $\pm$ 12.2)	3-20mm	Fragments and pellets	Tropical, North Pacific	Spear et al. 1995
<i>Puffinus griseus</i>	Sooty shearwater	218	88.5	/		Pellets	Offshore, North Pacific	Ogi et al. 1990

<i>Puffinus griseus</i>	Sooty shearwater	20	75	3.4		Pellets	Offshore eastern North Pacific	Blight and Burger 1997
<i>Puffinus griseus</i>	Sooty shearwater	50	72	2.48 ( $\pm$ 2.7)		Fragments and pellets	Nova Scotia, Canada	Bond et al. 2014
<i>Puffinus mauretanicus</i>	balaric shearwater?	46	70	2.5 ( $\pm$ 2.9)	3.5 ( $\pm$ 10.5 <sup>A</sup> )		Catalan coast, Mediterranean	Codina-García et al. 2013
<i>Puffinus nativitatis</i>	Christmas shearwater	5	40	1	3-5mm	Fragments and pellets	Tropical, North Pacific	Spear et al. 1995
<i>Puffinus pacificus</i>	Wedge-tailed shearwater	23	4	2.5 ( $\pm$ 2.1)		Fragments and pellets	Tropical, North Pacific	Spear et al. 1995
<i>Puffinus pacificus</i> dark phase	Wedge-tailed shearwater	62	24.2	3.5 ( $\pm$ 2.7)		Fragments and pellets	Tropical, North Pacific	Spear et al. 1995
<i>Puffinus pacificus</i>	Wedge-tailed shearwater	20	60	max: 11	Pellets 2-4mm	Pellets	Hawaii	Fry et al. 1987
<i>Puffinus puffinus</i>	Manx shearwater	10	30	0.4		Pellets	Rhum, UK	Furness 1985b
<i>Puffinus puffinus</i>	Manx shearwater	25	60	/	<10 mm		Rio Grande do Sul, Brazil	Colabuno et al. 2009
<i>Puffinus puffinus</i>	Manx shearwater	6	17	/		Fragments	Rio Grande do Sul, Brazil	Colabuno et al. 2010
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	6	100	19.8	1-4mm	Pellets	California, USA	Baltz and Morejohn 1976
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	324	81.8	/		Pellets	Offshore, North Pacific	Ogi et al. 1990
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	330	83.9	5.8 ( $\pm$ 0.4*)	2-5mm	Pellets	Bering Sea, North Pacific	Vlietstra and Parga 2002
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	5	80	/		Fragments and pellets	Alaska, USA	Robards et al. 1995
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	99	100	15.1 ( $\pm$ 13.2)	>2mm		Offshore, North Pacific	Yamashita et al. 2011
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	129	67	Adults: 4.5	0.97-80.8mm	Fragments	North Stradbroke Island, Australia	Acampora et al. 2013

		Juvenile: 7.1					
<i>Puffinus tenuirostris</i>	Short-tailed shearwater	12	100	27	>2mm	Offshore, North Pacific	Tanaka et al. 2013
<i>Puffinus yelkouan</i>	Yelkouan shearwater	31	71	4.9 ( $\pm$ 7.3)	4.0 ( $\pm$ 13.0 <sup>A</sup> )	Catalan coast, Mediterranean	Codina-García et al. 2013
Antarctic petrel ( <i>Thalassoica antarctica</i> )		184	< 1	/	Fragments, pellets 3-6mm	Antarctica	Ainley et al. 1990
<a href="#">Family Hydrobatidae</a>	-						
<i>Fregatta grallaria</i>	White-bellied storm petrel	13	38	1.2	Pellets Max mass: 0.042g	Gough Island, UK South Atlantic	Furness 1985a
<i>Fregatta grallaria</i>	White-bellied storm petrel	296	< 1	1	Fragment	Offshore, North Pacific	Spear et al. 1995
<i>Fregatta grallaria</i>	White-bellied storm petrel	318	/	1987-89 <sup>B</sup> 0.63 $\pm$ 1.13	Pellets 33.3%	Southern Ocean	Ryan 2008
<i>Fregatta grallaria</i>	White-bellied storm petrel	137	/	1999 <sup>B</sup> 0.63 $\pm$ 1.37	Pellets 20.9%	Southern Ocean	Ryan 2008
<i>Fregatta grallaria</i>	White-bellied storm petrel	95	/	2004 <sup>B</sup> 0.72 $\pm$ 1.87	Pellets 16.2%	Southern Ocean	Ryan 2008
<i>Garrodia nereis</i>	Grey-backed storm petrel	11	27	0.3	Pellets: Max mass: 0.010g	Gough Island, UK South Atlantic	Furness 1985a
<i>Garrodia nereis</i>	Grey-backed storm petrel	12	8.3	/	Pellets	Southern Ocean	Ryan 1987
<i>Oceanodroma furcata</i>	Fork-tailed storm petrel	/	/	/	<5mm	Aleutian Islands, USA	Ohlendorf et al. 1978
<i>Oceanodroma furcata</i>	Fork-tailed storm petrel	21	85.7	Max: 12	Pellets 22%	Alaska, USA	Robards et al. 1995
<i>Oceanodroma furcata</i>	Fork-tailed storm petrel	7	100	20.1	Pellets 16%	Eastern North Pacific	Blight and Burger 1997

<i>Oceanodroma leucorhoa</i>	Leach's storm petrel	15	40	1.66 ( $\pm$ 1.2)	2-5mm	Newfoundland, Canada	Rothstein 1973
<i>Oceanodroma leucorhoa</i>	Leach's storm petrel	17	58.8	2.9	Pellets	St. Kilda, Scotland, UK	Furness 1985b
<i>Oceanodroma leucorhoa</i>	Leach's storm petrel Leach's storm petrel	354	19.8	3.5 ( $\pm$ 2.6)	Fragments, pellets 2-5mm	Offshore, North Pacific	Spear et al. 1995
<i>Oceanodroma leucorhoa</i>	Leach's storm petrel	64	48.4	Max: 13	Monofilament line, fragments, pellets	Alaska, USA	Robards et al. 1995
<i>Oceanites oceanicus</i>	Wilson's storm petrel	20	75	4.4	2.9mm	Ardery Island, Antarctica	van Franeker and Bell 1988
<i>Oceanites oceanicus</i>	Wilson's storm petrel	91	19	/	Fragments, pellets 3-6mm	Antarctica	Ainley et al. 1990
<i>Oceanites oceanicus</i>	Wilson's storm petrel	133	38.3	Stomach = 1.4 Gizzard = 5.4	26% beads	North Carolina, USA	Moser and Lee 1992
<i>Pelagodroma marina</i>	White-faced storm petrel	19	84	11.7	Pellets Max mass: 0.34g	Gough Island, UK South Atlantic	Furness 1985a
<i>Pelagodroma marina</i>	White-faced storm petrel	15	73.3	13.2 $\pm$ 9.5	Pellets 2-5mm	Offshore, North Pacific	Spear et al. 1985
<i>Pelagodroma marina</i>	White-faced storm petrel	24	20.8	/	Pellets 41%	Southern Hemisphere	Ryan 1987
<i>Pelagodroma marina</i>	White-faced storm petrel	253		1987-89 <sup>B</sup> 3.98 $\pm$ 5.45	Pellets 69.6%	Southern Ocean	Ryan 2008
<i>Pelagodroma marina</i>	White-faced storm petrel	86	/	1999 <sup>B</sup> 4.06 $\pm$ 5.93	Pellets 37.5%	Southern Ocean	Ryan 2008
<i>Pelagodroma marina</i>	White-faced storm petrel	5	/	2004 <sup>B</sup> 2.52 $\pm$ 4.43	Pellets 13.5%	Southern Ocean	Ryan 2008
Family Diomedidae							

<i>Phoebastria fusca</i>	Sooty albatross	73	42.7	/	Pellets 34%	Southern Ocean	Ryan 1987
<i>Phoebastria immutabilis</i>	Laysan albatross	/	52	/	Pellets 2-5mm	Hawaiian Islands, USA	Sileo et al. 1990
<i>Phoebastria nigripes</i>	Black-footed albatross	/	12	/	Pellets 2-5mm	Hawaiian Islands, USA	Sileo et al. 1990
<i>Phoebastria nigripes</i> (As <i>Diomedea nigripes</i> )	Black-footed albatross	3	100	5.3	Pellets 50%	Offshore, eastern North Pacific	Blight and Burger 1997
<i>Thalassarche melanophri</i>	Black-browed albatross	2	100	3	Pellets 50%	Rio Grande do Sul, Brazil	Tourinho et al. 2010
<a href="#">Order Charadriiformes</a>							
<a href="#">Family Laridae</a>							
<i>Larus audouinii</i>	Audouin's gull	15	13	49.3 ( $\pm$ 77.7)	2.5 ( $\pm$ 5.0 <sup>*</sup> )	Catalan coast, Mediterranean	Codina-García et al. 2013
<i>Larus glaucescens</i>	Glaucous-winged gull	589 boluses	12.2	/	<10mm	Protection Island, USA	Lindborg et al. 2012
<i>Larus heermanni</i>	Heermann's Gull	15	7	1	Pellets 1-4mm	California, USA	Baltz and Morejohn 1976
<i>Larus melanocephalus</i>	Mediterranean gull	4	25	3.7 ( $\pm$ 7.5)	3.0 ( $\pm$ 5.0 <sup>*</sup> )	Catalan coast, Mediterranean	Codina-García et al. 2013
<i>Larus michahellis</i>	Yellow-legged gull	12	33	0.9 ( $\pm$ 1.5)	2.0 ( $\pm$ 8.0 <sup>*</sup> )	Catalan coast, Mediterranean	Codina-García et al. 2013
<i>Rissa brevirostris</i>	Red-legged kittiwake	15	26.7	/	Pellets: Mean 5.87mm	Alaska, USA	Robards et al. 1995
<i>Rissa tridactyla</i>	Black-legged kittiwake	8	8	4	Pellets 1-4mm	California, USA	Baltz and Morejohn 1976
<i>Rissa tridactyla</i>	Black-legged kittiwake	256	7.8	Max: 15	Pellets	Alaska, USA	Robards et al. 1995

<i>Rissa tridactyla</i>	Black-legged kittiwake	4	50	1.2 ( $\pm$ 1.9)	3.0 ( $\pm$ 5.0 <sup>*</sup> )	Catalan coast, Mediterranean	Codina-García et al. 2013
<a href="#">Family Alcidae</a>							
<i>Aethia psittacula</i>	Parakeet auklet	/	/	/	<5mm	Aleutians Islands, USA	Ohlendorf et al. 1978
<i>Aethia psittacula</i>	Parakeet auklet	208	93.8	17.1	Pellets 4.08mm	Alaska, USA	Robards et al. 1995
<i>Fratercula cirrhata</i>	Tufted puffin	489	24.5	Max: 51	Pellets 4.10mm	Alaska, USA	Robards et al. 1995
<i>Fratercula cirrhata</i>	Tufted puffin	9	89	3.3	Pellets	Offshore, North Pacific	Blight & Burger 1997
<i>Fratercula corniculata</i>	Horned puffin	/	/	/	<5mm	Aleutian Islands, USA	Ohlendorf et al. 1978
<i>Fratercula corniculata</i>	Horned puffin	120	36.7	Max: 14	Pellets 5.03mm	Alaska, USA	Robards et al. 1995
<i>Fratercula corniculata</i>	Horned puffin	2	50	1.5	Pellets	Offshore, North Pacific	Blight and Burger 1997
<i>Uria aalge</i>	Common murre	1	100	2011 – 2012 1	6.6 ( $\pm$ 2.2)	Newfoundland, Canada	Bond et al. 2013
<i>Uria lomvia</i>	Thick-billed murre	186	11	0.2 ( $\pm$ 0.8)	4.5 ( $\pm$ 3.8)	Canadian Arctic	Provencher et al. 2010
<i>Uria lomvia</i>	Thick-billed murre	3	100	2011 – 2012 1	6.6 ( $\pm$ 2.2)	Newfoundland, Canada	Bond et al. 2013
<i>Uria lomvia</i>	Thick-billed murre	1249	7.7	1985 – 1986 0.14 ( $\pm$ 0.7 <sup>*</sup> )	10.1 ( $\pm$ 7.4)	Newfoundland, Canada	Bond et al. 2013
<a href="#">Family Stercorariidae</a>							
<i>Stercorarius antarcticus</i> (as <i>Catharacta antarcticu</i> )	Brown skua	494	22.7	/	Pellets 67%	Southern Ocean	Ryan 1987
<i>Stercorarius hamiltoni</i> (as <i>Catharacta</i> )	Tristan skua	11	9	0.3	Pellets	Gough Island, UK	Furness 1985a



<i>hamiltoni</i> )				Max: 3	Max mass: 0.064g	South Atlantic	
<i>Stercorarius longicaudus</i>	Long-tailed skua	2	50	5	Fragments, pellets	Eastern North Pacific	Spear et al. 1995
<i>Stercorarius parasiticus</i> )	Arctic skua	2	50	/	Pellets 50%	Southern Ocean	Ryan 1987
<u>Family Scolopacidae</u>							
<i>Phalaropus fulicarius</i>	Grey phalarope	20	100	Max: 36	Beads 1.7-4.4mm	California, USA	Bond 1971
<i>Phalaropus fulicarius</i>	Grey phalarope	7	85.7	5.7	Pellets	California, USA	Connors and Smith 1982
<i>Phalaropus fulicarius</i>	Grey phalarope	2	50	/	Pellets	Southern Ocean	Ryan 1987
<i>Phalaropus fulicarius</i>	Grey phalarope	55	69.1	Stomach = 1 Gizzard = 6.7	Beads 16.7%	North Carolina, USA	Moser and Lee 1992
<i>Phalaropus lobatus</i> )	Red-necked phalarope	36	19.4	Stomach = 0 Gizzard = 3.7	Beads 16.7%	North Carolina, USA	Moser and Lee 1992
<u>Family Sternidae</u>							
<i>Onychoprion fuscatus</i>	Sooty tern	64	1.6	2	Pellets 4mm	Offshore, eastern North Pacific	Spear et al. 1995
<i>Gygis alba</i>	White tern	8	12.5	5	Fragments 3-4mm	Offshore, eastern North Pacific	Spear et al. 1995
<u>Order Suliformes</u>							
<u>Family Phalacrocoracidae</u>							
<i>Phalacrocorax atriceps purpurascens</i>	Macquarie shag	<sup>c</sup> 64	7.8	1 per bolus	Polystyrene spheres	Macquarie Island, Australia	Slip et al. 1990

## Annex VII. Estimated cost of marine litter for the EU fishery sector

Table VII.1. Estimated cost of marine litter for the EU fishery sector (based on Mouat et al. 2010 in Arcadis 2014)

Type of cost	Cost per vessel (€)	Estimated cost for the EU (M€)	Calculation method
Reduced catch revenues (contamination forces fishermen to use more time for the selection of their catches and to discard part of them)	2,340	28.64	The cost estimated by Mouat et al. (2010) for Scottish vessels (€2,200 per vessel per year), actualised in 2013 prices, was multiplied by the number of EU trawlers (EU vessels that use seafloor fishing gear), i.e. 12,238.
Removing litter from fishing gear	959	11.74	The time needed to remove litter from fishing gear, as estimated by Mouat et al (2010) for Scottish vessels (41 hours per vessel per year), was multiplied by the average EU27 labour cost (€23.4 per hour) and then by the number of EU trawlers (EU vessels that use seafloor fishing gear), i.e. 12,238.
Broken gear, fouled propellers	191	16.79	The cost related to broken gear and fouled propellers, as estimated by Mouat et al. (2010) for Scottish vessels (€180 per vessel per year), actualised in 2013 prices was multiplied by the total number of fishing vessels in the EU (87,667 according to Eurostat).
Cost of rescue services	52	4.54	The average cost of incidents around the British Isles attended by the Royal National Lifeboat Institution (RNLI) in 1998 (£4,000 per vessel) was multiplied by the number of incidents (200), and divided by the number of UK fishing boats (7,800), as indicated by Fanshawe (2002). The estimated yearly cost per boat resulting by this calculation was then multiplied by 31.1%, i.e. the share of rescue operation dedicated to fishing vessels, as indicated for the UK by Mouat et al (2010) (year 2008). The result (£32 per vessel) was then actualised in 2013 prices and converted to € and multiplied by the total number of fishing vessels in the EU (87,667 according to Eurostat).
<b>Total</b>		<b>61.71</b>	

Annex VIII. Estimated clean-up and management costs of marine litter  
 Table VIII.1 Estimated clean-up and management costs of marine litter – some examples

Country / Region	Estimated cost at national and municipality level	Source
Belgium and Netherlands	USD 13.8 million (EUR 10.4 million) for all municipalities in Belgium and Netherlands (ave. USD 264 885/municipality/year (EUR 200 000/municipality/ year; EUR 629 – 97 346 per km)) Costs are higher for areas with high visitor numbers; for example the Den Haag Municipality spends USD 1.43 million/year (EUR 1.27 million/year) with costs for processing litter (including transport) about USD 229/tonne (EUR 165/tonne).	Mouat et al, 2010 OSPAR 2009
Peru	USD 2.5 million in labour costs (ave. USD 400 000/year in municipality of Ventanillas)	Alfaro, 2006 cited in UNEP, 2009
UK	USD 24 million (EUR 18 million) (ave. USD 193 365/municipality/year (EUR 146,000/municipality/ year) (per km cleaning costs range from USD 226-108 600/km/year (EUR 171-82 000/km/year)). Specific municipality costs: <ul style="list-style-type: none"> <li>• Suffolk: approx. USD 93 500/year (GBP 60 000/year) on 40km of beaches</li> <li>• Carrick District Council (Devon): approx. USD 56 000/year (GBP 32 000/year) on 5km of beaches.</li> <li>• Studland (Dorset): USD 54 000/year (GBP 36,000/year) to collect 12-13 tonnes of litter each week in the summer along 6km of beaches.</li> <li>• Kent coastline: direct and indirect cost of litter estimated at over USD 17 million/year (GBP 11 million/year).</li> <li>• Annual expenditure on beach cleaning in 56 local authorities ranged from USD 23/km (GBP 15/km) in West Dunbartonshire to USD 78,000/km (GBP 50 000/km) in Wyre.</li> </ul>	Mouat et al, 2010 Fanshawe and Everard, 2002 OSPAR 2009
Bay of Biscay and Iberian coast	A Spanish council with 30 beaches (5 Blue Flags) spends around USD 111 000/year (EUR 80 000/year) on beach cleaning  A French council with 30 beaches (5 Blue Flags) spends around USD 556 000/year (EUR 400 000/year) on 'beach caring' (including beach clearing, monitoring of buoys, coastguards etc.), of which around 20% (USD 111 000 (EUR 80 000)) relates to beach clearing.  In Landes, the cost of cleaning up 108km of sandy beaches was USD 11 million (EUR 8 million) between 1998 and 2005  Cost of beach cleaning between USD 6 250-69 460/year/council (EUR 4 500-50 000/year/council) corresponding to average cost of USD 9 000/km (EUR 6 500/km) of cleaned beach/year.	OSPAR, 2009
Poland	Beach cleaning and removing litter from harbour waters cost USD 792 000 (EUR 570 000) in 2006 (same amount also spent in five communes and two ports)	(UNEP, 2009)
Oregon, California, Washington (USA)	Annual combined expenditure of USD 520 million (USD 13/resident/year) to combat litter and curtail potential marine litter	Stickel et al., 2012
APEC region	USD 1 500/tonne of marine litter in 2007 terms	(McIlgorm, 2009)

Annex IX Compilation of Eleven Best Practices in European Seas (evaluated using the DeCyDe-4-MARLISCO tool)

	<b>Title</b>	<b>Implementation area</b>	<b>Implementation scale</b>	<b>Duration (y)</b>	<b>Theme(s)</b>	<b>Type of initiative<sup>1</sup></b>
1	Operation clean coasts 'Calanques Propres'	France	Sub-national	>5	Mitigation Awareness	Campaign P-A-A
2	Responsible snack bar project	Spain	National	0-1	Prevention	Econ./Market instrument
3	Sea surface marine litter cleaning operation	Turkey	Sub-national	>5	Mitigation	P-A-A
4	Integrated action plan for the cleaning of the Channel coast	France	Sub-national	>5	Prevention Mitigation Awareness	P-A-A campaign
5	The plastic bag levy	Ireland	National	>5	Prevention	Policy/Reg. Impl. Econ./Market instrument
6	Coastwatch Portugal campaign	Portugal	National	>5	Mitigation Awareness	Campaign
7	Fishing for litter	Netherlands	Sub-national	2-5	Mitigation Awareness	P-A-A
8	Blue lid campaign	Turkey	National	1-2	Awareness	P-A-A campaign
9	Separation and recycling of materials from fishing nets and trawls	Denmark	National	>5	Prevention Mitigation	P-A-A other
10	BREF – best available techniques reference document – in common wastewater and waste gas treatment/management systems in the chemical sector	Europe	European	>5	Prevention	Policy/Reg. Impl.
11	Dive against debris, project AWARE	Global	Global	>5	Mitigation	P-A-A campaign

<sup>1</sup> Key to type of initiative: P-A-A – Practice/Activity/Action; Policy/Reg. Impl. – policy/regulation implementation; Econ./Market Instrument – economic and market-based instruments.

## Annex X. Sampling and analysis techniques for microplastics

### *Microplastics in Sediments*

A wide range of sampling techniques are used for monitoring microplastics in sediments reviewed in Hidalgo-Ruz et al. (2012), van Cauwenberghe et al. (2013) and Rocha-Santos and Duarte (2015). These methods include density separation, filtration and/or sieving (Hidalgo-Ruz et al. 2012, Rocha-Santos and Duarte 2015). Also, to facilitate the plastic extraction among organics components such as organic debris (shell fragments, small organisms, algae or sea grasses, etc.) and other items such as pieces of tar, other methods can be applied, such as enzymatic, CCL<sub>4</sub> or H<sub>2</sub>O<sub>2</sub> digestion of organic materials have been proposed (Galgani et al. 2011, Hidalgo-Ruz et al. 2012, Cole et al. 2014) such as for water samples.

The most common approach is to extract plastic particles from the sediment using a density separation based on the differences in density between plastic and sediment particles. Typically, this is achieved by agitating the sediment sample in concentrated sodium chloride (NaCl) solution. However, as the density of the NaCl solution is only 1.2 g cm<sup>3</sup>, only low density plastics will float to the surface and can hence be extracted. Different authors have addressed this issue by using different salt solutions to obtain higher densities. Corcoran et al. (2009) used a 1.4 g cm<sup>-3</sup> polytungstate solution, Imhof et al. (2013) extracted microplastics from sediments using zinc chloride (ZnCl<sub>2</sub>, 1.5-1.7 g cm<sup>3</sup>), while others (Dekiff et al. 2014, Van Cauwenberghe et al. 2013a, Van Cauwenberghe et al. 2013b) used a sodium iodide (NaI, 1.6 -1.8 g cm<sup>3</sup>) solution. These modifications result in an increased extraction efficiency for high density microplastics such as polyvinylchloride (PVC, density 1.14 - 1.56 g cm<sup>3</sup>) or polyethylene terephthalate (PET, density 1.32-1.41 g cm<sup>3</sup>). As these high-density plastics make up over 17% of the global plastic demand (PlasticsEurope 2013), not including these types of microplastics can result in a considerable underestimation of microplastic abundances in sediments. Especially as these high-density plastics are the first to settle and incorporate into marine sediments.

Sieves used in separation of particles usually have mesh sizes ranging from 38µm to 5 mm and often include 330µm, 1mm and 2mm. To avoid degradation, plastics separated from the sample have been dried and kept in the dark, however this step is probably unnecessary if samples are examined within a few months of collection.

Visual examination is the most common method to assess size and quantities of microplastics. Various imaging approaches, such as zooscan™ (Gilfillan 2009) or semi-automated methods (flow/cytometer, cell sorter, coulter counters) may be practical for the visualization or counting of microplastic particles, with the potential to enable a large number of samples to be analysed. For a better identification of plastics, specific criteria can be applied, such as the presence of cellular or organic structures, the constant thickness of fragments or fibres, homogeneous colours and plastic brightness. However, the reliability of such approaches has not been evaluated. Other analyses based on visual examination with light, polarised or not, or electron microscopy, may provide higher resolution but cannot be used to determine polymer type.

The choice of sampling strategy and sampling approach (reviewed by (Hidalgo-Ruz et al. 2012) will eventually determine the unit in which observed abundances will be reported. While a simple conversion can sometimes be made to compare among studies (Lusher et al. 2015), comparison is often impossible or requires assumptions that lead to biased results. Studies sampling an area (using quadrants) will often report abundances per unit of surface (m<sup>-2</sup>); e.g. (Martins and Sobral 2011). If real bulk samples up to a specific depth are taken the reporting unit is m<sup>3</sup> (e.g. (Turra, et al. 2014)). Conversion between these types of abundances is possible, if sufficient information is available on sampling depth. Yet, for 20% of the studies this is not the case as reported sampling depths can range from 0 to 50 cm. Other widely used reporting units are volume (mL to L; e.g. Noren 2007) or weight (g to kg; e.g. Claessens et al., 2011, Ng and Obbard 2006). Conversion between these two types of units is not straight forward. Detailed information on the density of the sediment is required. As this is never (as far as we could establish) reported in microplastic studies, assumptions have to be made. For example, the conversion of microplastic abundances in sediment (Claessens et al. 2011). Additionally, within studies reporting weight, a distinction can be made among those reporting wet (sediment) weight and those reporting dry weight. This adds to the constraints of converting from weight to volume units, or vice versa. Sediment samples from different locations or even different zones on one beach have different water content. Therefore, a (limited) number of authors choose to express microplastic abundance per sediment as dry weight to eliminate this variable (Claessens et al. 2013, Dekiff et al. 2014, Ng and Obbard 2006, Van Cauwenberghe et al. 2013); (Vianello et al. 2013).

### *Microplastics in Biota*

In terms of monitoring and with regards to "in situ" experiments, one of the most important aspects is the choice of target species. It is important to consider (i) the exposure to plastics, especially for the species that are living at the surface or in the sediments, (ii) the ingestion rate, especially for filter feeders such as bivalves, (iii) the significance of results which vary depending on whether environmental impact or human health is considered, (iv) the biological sensitivity of certain species, such as the high retention rate in birds of the procellariiform family, and (v) a large distribution and easy sampling of the target species.

Biological sampling that involves the examination and characterisation of plastic fragments consumed by marine organisms has been used for fishes (Lusher et al. 2013, Choy and Drazen 2013, Avio, Gorbi et al. 2015), invertebrates (Browne et al. 2008, Murray and Cowie 2011, Desforges et al. 2015, Van Cauwenberghe et al., 2015) and birds ((van Franeker et al. 2011). In general, the research question addressed will greatly influence which sampling and extraction technique to use. For example, size range of microplastics can be related to the micro- and macro-plankton highlighting the potential for microplastic ingestion by a wide variety of organisms (Hidalgo-Ruz et al. 2012). Thus, the sampling scale and methodology will depend on the size of the particle or the size group of the studied organisms. However, harmonisation of sampling and extraction techniques should be adopted for monitoring purposes.

The methodological difficulties in isolation protocols partly explain why only a few studies specifically addressed the occurrence of microplastics in marine organisms. Even though suitable methods have been identified for sediment and water samples, the extraction and quantification of microplastics from organisms may be masked within biological material and tissues. Protocols on the extraction of microplastics from marine invertebrates after a pre-digestion of organic matter have been proposed (Claessens, Van Cauwenberghe et al. 2013), indicating the importance of solvent properties and pH for sample treatment, affecting both the estimation and the characterization of the polymers by FT-IR. The enzymatic digestion of organic matter with proteinase k is a reliable method to extract microplastics from planktons samples (Cole et al. 2014), but at higher costs when considering large scale field sampling and monitoring.

Annex XI. Revised GPML Indicators and Targets  
 Indicators and targets - GPML implementation & related processes

Table 1. Generic Indicators – Goals A, B and C

<b>Intended Outcome</b>	<b>Indicator of GPML Implementation</b>	<b>Target (by December2016)<sup>2</sup></b>	<b>Monitoring/Verification</b>
Operational partnership with a wide range of partners facilitated through an online forum promoting the Honolulu Commitment and Strategy	Number of Governments, organisations, agencies and institutions joining the GPML.	>100	Number of submitted forms to join the GPML
	An effective and functional international steering committee (SC)	SC established according to Terms of Reference and meeting at least once per year	SC meeting report, containing clear guidance to develop the GPML
	An effective and functional set of Regional Nodes	Four Regional Nodes established according to Terms of Reference with developed networks operational	Regional Nodes report to GPML Secretariat and Focal Areas A, B and C
	Meeting of the global partnership to review implementation of the Honolulu Strategy	Partnership meeting	Meeting report with recommendations for improving implementation of the GPML and associated management measures
Development of regional and national policy instruments aligned with the 'Honolulu Strategy'	Number of regional <sup>3</sup> and national policy instruments aligned with the Honolulu Strategy discussions for decision-making at respective levels.	5 regional policy instruments 10 national policy instruments	Policy instruments

<sup>2</sup>December 2016 is initial target date. Further targets to be agreed as the GPML develops.

<sup>3</sup>Regional in this context refers to multi-national bodies, agreements and other arrangements, such as Regional Seas Organisations. In some countries, regional is used to indicate sub-national levels of governance or organisation.

Table 2. Indicators and Targets - GPML Outputs

<b>Intended outcome</b>	<b>Indicator of GPML outputs</b>	<b>Target (by December 2016)</b>	<b>Monitoring/verification</b>
Operational partnership promoting the GPML Honolulu Strategy by the production of reports, articles, videos, training materials and related products and activities	Number of activities	1 per Region	Report uploaded to MLN
	Production of Steering Committee reports	1 per year from each	Reports approved by GPML Secretariat
	Production of GPML Newsletter/webinar	At least annual	Newsletter produced by GPML Secretariat
	Demonstration Project progress reports	1 annual progress report per project	Reports approved by GPML Secretariat



Table 3. Indicators and Targets - Demonstration Projects

Specific Land-Based Indicators based on Demonstration Projects – Goals A and C

<b>Intended outcome</b>	<b>Indicator description</b>	<b>Target (by 2020)<sup>4</sup></b>	<b>Monitoring/verification</b>
Reduction of influx of solid waste to the marine environment through the demonstration of good policy and on-the-ground practices and technologies, including the introduction of new instruments and market-based policies	Reduction in the direct entry of plastic to the marine environment by improved waste management	20% reduction in marine input in 5 demonstration projects <sup>5</sup>	Self-reporting & project reports Independent assessment of degree of reduction of inputs and cost-benefit analysis.
	Increase in recycling rates of specified wastes	50% increase in recycling rates in 5 demonstration projects	Self-reporting & project reports Independent assessment of degree of increase of recycling
	Reduction in demand for ‘single-use’ plastic shopping bags <sup>6</sup>	25% reduction in demand in 5 countries	National reporting
	Agreement to adopt new good practises resulting from demonstration projects	10 Governments or private sector organisations agree to make use of good practises <sup>7</sup>	Self-reporting of proposed actions
	Number of illegal waste dumps on coast	Significant reduction <sup>8</sup>	National reporting

<sup>4</sup>Dependent on: i) the timescale for introduction of demonstration projects and other measures; ii) the scale and complexity of the socio-ecological system; iii) the willingness of all relevant stakeholders to play an active role; iv) the availability of technical know-how and funding as required; and, v) any in-built hysteresis in the social, economic, physical or ecological elements of the system (Oosterhuis et al. 2014).

<sup>5</sup>To include representative sectors, for example: illegal waste dumps, coastal tourism, waste management in urban areas, retail sector and Small Island Developing States (SIDS).

<sup>6</sup>For example, by introducing a charge per bag and encouraging more durable multiple-use replacements

<sup>7</sup>To include representative sectors, for example: illegal waste dumps, coastal tourism, waste management in urban areas, retail sector and SIDS.

<sup>8</sup>‘Significant reduction’ – this will be dependent on a number of factors including the chain of responsibility, context, identifying manageable sources and the cost-benefit of introducing reduction measures

Table 4. Specific Sea-based Indicators based on Demonstration Projects – Goals B and C

<b>Intended Outcome</b>	<b>Indicator Description</b>	<b>Target (2020)<sup>9</sup></b>	<b>Monitoring/Verification</b>
Reduction of influx of solid waste to the marine environment through the demonstration of good policy and on-the- ground practices and technologies, including the introduction of new instruments and market-based policies	Reduction in the direct entry of plastic to the marine environment by improved waste management	20% reduction in marine input in 5 demonstration projects <sup>10</sup>	Self-reporting & project reports Independent assessment of degree of reduction of inputs and cost-benefit analysis.
	Increase in recycling rates of specified wastes	50% increase in recycling rates in 5 demonstration projects	Self-reporting & project reports Independent assessment of degree of increase of recycling
	Agreement to adopt new good practices resulting from demonstration projects	10 Governments or private sector organisations agree to make use of good practices <sup>11</sup>	Self-reporting of proposed actions

<sup>9</sup>Dependent on: i) the timescale for introduction of demonstration projects and other measures; ii) the scale and complexity of the socio-ecological system; iii) the willingness of all relevant stakeholders to play an active role; iv) the availability of technical know-how and funding as required; and v) any in-built hysteresis in the social, economic, physical or ecological elements of the system.

<sup>10</sup>To include representative sectors, for example: aquaculture, fisheries, shipping, cruise industry and recreational boating.

<sup>11</sup>To include representative sectors, for example: aquaculture, fisheries, shipping, cruise industry and recreational boating.

Table 5. Indicators and Potential Targets - Environmental State<sup>12</sup> - Goals A, B and C

Generic Indicators – Goal C

<b>Intended outcome</b>	<b>Indicator description</b>	<b>Target (2020-25)</b>	<b>Monitoring/verification</b>
Reduce the quantities and impact on the environment of marine litter entering from all sources	Number of cetaceans injured or killed	Significant reduction <sup>13</sup>	IWC, Regional Seas Bodies, national government, municipalities and NGO reporting
	Number of turtles killed by entanglement	Significant reduction	Regional Seas Bodies, national government, municipalities and NGO reporting
	Quantity of plastic (number and mass of items) in guts of indicator species from necropsies (e.g. fish, birds, reptiles, cetaceans)	Significant reduction	Regional Seas Bodies, national government, municipalities and NGO reporting
	Number and mass of items of floating macro-litter (items km <sup>-2</sup> )	Significant reduction	Regional Seas Bodies, national government and NGO reporting
	Number of items of floating micro-litter, especially microplastics (items km <sup>-2</sup> )	Significant reduction	Regional Seas Bodies, national government and NGO reporting
	Number and mass of items of litter on shorelines - km <sup>-1</sup> shoreline		Regional Seas Bodies, national government and NGO reporting

Table 6. Specific Land-based Indicators – Goals A and C

<b>Intended Outcome</b>	<b>Indicator description</b>	<b>Target (2020-25)</b>	<b>Monitoring/verification</b>
Reduce the quantities and impact on the environment of marine litter introduced on land and entering the sea	Quantity of litter on tourist beaches - km <sup>-1</sup> shoreline	Significant reduction <sup>14</sup>	Regional Seas Bodies, national government, municipalities and NGO reporting

<sup>12</sup>See JRC/EC 2013 for a comprehensive description of potential indicators of marine litter

<sup>13</sup> ‘Significant reduction’ – this will be dependent on a number of factors including the chain of responsibility, context, identifying manageable sources and the cost-benefit of introducing reduction measures

<sup>14</sup> ‘Significant reduction’ – this will be dependent on a number of factors including the chain of responsibility, context, identifying manageable sources and the cost-benefit of introducing reduction measures

– Goals B and C

<b>Intended Outcome</b>	<b>Indicator Description</b>	<b>Target (2020-25)</b>	<b>Monitoring/Verification</b>
Reduce the quantities and impact on the environment of marine litter introduced directly at sea	Quantity (volume m <sup>3</sup> and length km) of capture fisheries gear abandoned, lost or otherwise discarded (ALDFG) (e.g. nets, lines, pots, FADs)	Significant reduction <sup>15</sup>	FAO reporting (LC/LP), Regional Seas Bodies, national governments, municipalities, fisheries industry
	Quantity of other capture fisheries-related items in the environment – items km <sup>-2</sup> sea surface, km <sup>-2</sup> water column, km <sup>-2</sup> seabed, km <sup>-1</sup> shoreline (e.g. strapping bands, boxes, rope)	Significant reduction	Reporting by NGOs, Regional Seas Bodies, national governments, municipalities, fisheries industry
	Quantity (volume m <sup>3</sup> and length km) of aquaculture gear abandoned, lost or otherwise discarded (ALDFG) - items km <sup>-2</sup> sea surface, km <sup>-2</sup> water column, km <sup>-2</sup> seabed, km <sup>-1</sup> shoreline (e.g. floats, rope, nets, cages, poles)	Significant reduction	FAO reporting; regional reporting e.g. Network of Aquaculture Centres in Asia-Pacific (NACA), NGOs, Regional Seas Bodies, national governments, municipalities
	Quantity of litter derived from commercial shipping	Significant reduction	National governments, NGOs, Regional Seas Bodies & municipalities reporting
	Quantity of litter derived from cruise industry	Significant reduction	National reporting
	Number of turtles killed by ALDFG	Significant reduction	CBD, Regional Seas Bodies, national and NGO reporting
	Number of cetaceans injured by ALDFG	Significant reduction	FAO, IWC, CBD, Regional Seas Bodies, national and NGO reporting
	Number of fish killed by ALDFG	Significant reduction	FAO, CBD, Regional Seas Bodies, national and NGO reporting
	Number of birds killed by ALDFG	Significant reduction	CBD, Regional Seas Bodies, national and NGO reporting
	Number of containers and other cargo lost by commercial shipping	Significant reduction	National and shipping industry reporting

<sup>15</sup>Significant reduction’ – this will be dependent on a number of factors including the chain of responsibility, context, identifying manageable sources and the cost-benefit of introducing reduction measures

Indicators of Social and Economic Impacts - Goal C

<b>Intended Outcome</b>	<b>Indicator Description</b>	<b>Target (2020-25)</b>	<b>Monitoring/Verification</b>
Reduce the social and economic impact on the environment of marine litter entering from all sources	Number of vessels damaged or lost due to collisions or entanglement (e.g. fouled propellers or blocked cooling water intake)	Significant reduction <sup>16</sup>	Operators, national governments
	Loss of energy generation capacity (and income) and risk of accidental damage due to blocked cooling water intakes in coastal power stations, including nuclear power stations; loss of functioning of desalination plants.	Significant reduction	Operators, national governments
	Cost of beach cleaning	Significant reduction	Municipalities
	Number of injuries to public caused by marine litter	Significant reduction	National governments, municipalities, health authorities
	Number of call-outs of emergency services by stricken vessels	Significant reduction	National governments, emergency services, municipalities

<sup>16</sup> ‘Significant reduction’ – this will be dependent on a number of factors including the chain of responsibility, context, identifying manageable sources and the cost-benefit of introducing reduction measures

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